

## Journal of Science & Cycling Breakthroughs in Cycling & Triathlon Sciences

**Review Article** 

# Methodological Approaches in Testing Maximal Lactate Accumulation Rate - vLamax: A Systematic Review

Jamie Langley <sup>1</sup> , Ralf Haase <sup>2</sup> , Nico Nitzsche <sup>2</sup> and Michael Porter <sup>3</sup>

**Received:** 6 January 2025 **Accepted:** 6 August 2025 **Published:** 10 September 2025

- Department of Higher Education Sport, Loughborough College, Loughborough, UK.
- <sup>2</sup> Department of Sports Medicine and Exercise Therapy, Chemnitz University of Technology, Chemnitz, Germany.
- <sup>3</sup> Centre for Physical activity, and life sciences, University of Northampton, Northampton, UK.

#### Correspondence

Jamie Langley

Department of Higher Education Sport, Loughborough College, Loughborough, UK.

jamie.langley@loucoll.ac.uk

#### **Abstract**

In 1984 Mader constructed a mathematical model of human energy metabolism to understand the metabolic origin behind the maximal lactate steady state. An integral parameter of Mader's model requires knowledge of the maximal rate of glycolysis, which Mader derived from the maximal lactate formation rate within the muscle cell. However, in-vivo the maximal lactate formation rate within the muscle cannot be measured. Subsequently, Mader proposed the utility of measuring the rate of maximal blood lactate accumulation following supramaximal exercise as an indirect measure of glycolytic flux, termed vLamax. Recently, the vLamax has gained popularity amongst researchers and practitioners as an indirect assessment method to determine the maximal glycolytic rate. Currently, there is a distinct lack of continuity in methodological approaches between researchers. Therefore, the primary aim of this systematic review was to evaluate the current methodological approaches applied to test the vLamax. Based on the findings we make practical recommendations for researchers to adopt to promote standardisation of test procedures. Comprehensive searches of the databases; PubMed, SCOPUS, Google Scholar, and SPOLIT, identified 3545 articles for screening (1984-2024). In total 27 articles were included within this review, with seven different modalities identified. The results from this systematic review highlight several key considerations which need to be considered when testing the vLamax including; alactic timespan, test duration, baseline blood lactate concentration, modality specificity, movement velocity, recovery procedures, and post exercise blood lactate sampling times. Based on these findings this review provides detailed recommendations to standardise vLamax methods considering pre-test, test, and post-test factors.

#### Keywords

Glycolysis;  $\nu$ Lamax; Maximal Lactate Accumulation; Anaerobic Power; Sprint Performance; Blood Lactate.



This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/) which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



### 1 Introduction

The performance of high intensity exercise is largely influenced by the maximal rate of the energy generating pathways to produce contractile force (Heck et al., 2003). A noninvasive identification of each energy system utilisation is widely sought after, as the detection of deficiencies, provide the basis for training recommendations. Direct measurements of the oxidative energy system can be analysed via expired air during exercise (Van Hooren et al., 2024). Whereas a direct measurement of alactic glycolytic energy capacity/power is challenging due to metabolic substrates located within the muscle cell (Heck et al., 2003). Direct measurements of muscle phosphates require complex and invasive procedures; muscle biopsy (Parolin et al., 1999), and phosphorus magnetic resonance spectroscopy (Wackerhage et al., 1998). Whilst alactic substrate utilisation can be estimated indirectly via post exercise expired air (Knuttgen, 1970) and near infrared spectroscopy (McCully et al., 1994). Currently, there is no noninvasive gold standard to determine glycolytic metabolic power.

Analysis of post exercise capillary blood lactate concentration to determine the maximal rate of blood lactate accumulation (vLamax), has gained traction as an indirect marker of the glycolytic rate. This approach has been applied across various exercise modalities following short-term maximal exercise; cycling (Hauser et al., 2014; Adam et al., 2015; Dunst et al., 2023a; Harnish et al., 2023; Yang et al., 2023; Haase et al., 2024; Langley et al., 2024; Meixner et al., 2024), hand-cycling (Quittmann et al., 2018; Quittmann et al., 2021a), running (Hanon et al., 2011; Wawer et al., 2020; Quittmann et al., 2020; 2021b; 2023), rowing (Held et al., 2023; Schunemann et al., 2023), swimming (Sperlich et al., 2010; Teixeira et al., 2022; Mavroudi et al., 2023), and following isokinetic force tests of maximal strength loads (Nitzsche et al., 2018a; Nitzsche et al., 2020).

The glycolytic power of an athlete is traditionally associated with accelerations and the sprint finish during endurance races (Quittmann et al., 2023). However, the interaction of the maximal glycolytic (vLamax) and oxidative power (VO<sub>2max</sub>), have been successfully applied mathematical to simulations of human energy metabolism, to predict the power at maximal lactate steady state (MLSS) (Mader, 1984; Mader and Heck, 1986; Mader, 2003; Hauser et al., 2014; Hommel et al., 2019; Poffé et al., 2024). The goldstandard for determining the MLSS requires multiple constant load trials, which are time consuming, training restrictive, and does not explain the underlying physiological mechanisms (Quittmann et al., 2020). Therefore, approaches applying the metabolic power from the vLamax and VO<sub>2max</sub> to calculate the MLSS provide the metabolic profile of the athlete in a time effective method. Subsequent insight may allow training interventions to be tailored to an individual's metabolic profile by altering either the athletes vLamax or VO2max to improve MLSS (Hommel et al., 2019).

### 1.1 Theoretical Concept of the Maximal Lactate Accumulation Rate (vLamax)

Lactate is an intermediate metabolite between glycolysis and mitochondrial respiration. Pyruvate is converted into lactate via the enzyme lactate dehydrogenase (LDH) at the end of glycolysis (Emhoff & Messonnier, 2023). The transportation of lactate between cells is intrinsically linked to the metabolic demand, with the production rate being dependent upon the rate of glycolytic activity (Rogatzki et al., 2015). Maximal muscular is contraction strongly glycolytically disturbs which metabolic activating, homeostasis, thus increasing the lactate efflux into other tissues (blood plasma, oxidative muscle fibres, and central organs) due to the increase in lactate and H+ concentrations (Brooks et al., 1999).

Early studies described the utilisation of blood glycolysis based on lactate concentrations due to increasing metabolic demand (e.g., Margaria et al., 1933, Margaria et al., 1971; Jacobs et al., 1983), as well as the time constants of lactate distribution between the compartments (Zouloumian & Freund, 1981; Freund et al., 1984). However, quantification of metabolic energy derived from lactate accumulation in the blood is controversial, with several authors contesting the validity (Ferretti, 2015). This has stunted the progress of knowledge regarding the energetics of supramaximal exercise (Ferretti, 2015). Whilst significant lactate production occurs during steady state exercise, the majority is oxidised, therefore the VO2 accurately represents the energy contribution of 'aerobic glycolysis' (Brooks, 2012). Consequently, the net rise in the body lactate pooling represents the energy of non-oxidative glycolysis following supramaximal exercise (Brooks, 2012). Due to the rapid dissociation of lactate with total body water following maximal exercise the rise in blood lactate accumulation can be used to estimate non-oxidative glycolysis (Brooks, 2012).

Due to limited ability to store ATP (~5-8 mmol per kg of muscle) (Harris et al., 1974; Dawson, 1983), ATP must continually be resynthesised by the three energy systems. Applying specific rates of reactions and the concentrations of metabolites dependent upon; the individuals muscle mass, vLamax, and oxidative capacity, Mader (1984; 2003; Mader & Heck, 1986) devised a mathematical model of human energy metabolism to understand the metabolic origin behind the MLSS. This systematic review will focus specifically on the vLamax constructed by Mader (1984; Mader & Heck 1986) to denote the maximal glycolytic rate of the muscle. A recent overview and simplification of Mader's (1984) mathematical model of human energy metabolism has been provided by Wackerhage et al. (2022). Mader's (1984; 2003; Mader & Heck, 1986) model suggests the vLamax as an important parameter in endurance performance, illustrating an inverse relationship between vLamax and the MLSS.

### 1.2 Development of Mader's Maximal Lactate Accumulation Formula

Mader first proposed the maximal lactate formation rate as part of his mathematical model of human energy metabolism to identify the maximal rate of glycolysis within the muscle (Mader, 1984; Mader and Heck 1986; Mader, 2003). Mader (1984; Mader and Heck 1986) calculated the rate of ATP resynthesis via glycolysis using two key principles: 1) That an increased concentration of ADP (Adenosine and **AMP** (Adenosine diphosphate) monophosphate) activate the upregulation of glycolysis, whereas a reduction in pH inhibits phosphofructokinase (PFK) activity (Dobson et al., 1986), thus slowing down the glycolytic rate (Mader and Heck, 1986; Mader, 2003). 2) Lactate is always the product of glycolysis due to the near-equilibrium reaction of lactate dehydrogenase (LDH) (Sahlin et al., 1976; Rogatzki et al., 2015). These principles allowed Mader (1984; Mader & Heck, 1986) to calculate the maximum rate of glycolysis based on the maximum rate of lactate formation within the muscle cell (equation 1):

Maximum lactate formation rate = 
$$\frac{dLa}{dt}$$

### Equation (1)

The maximal lactate formation rate within the muscle cannot be measured directly. Therefore, Mader (1994) proposed the utility of measuring blood lactate prior and post an allout sprint test as an indirect measure of the maximal glycolytic flux (equation 2). Later, mathematical modelling from Heck et al. (2003) proposed an optimal test duration of 10 s due to the suppression of PFK activity with increasing acidosis the greater the exercise

duration. It should be noted the vLamax calculated from changes in peripheral blood lactate concentration, can only provide an estimate of the muscles maximal glycolytic flux, due to the complex dynamics of diffusion from the active muscle cell and elimination processes (Mader, 1994). Due to their interdependence the exact contribution of the alactic and glycolytic energy metabolism cannot be determined and is influenced by the intensity and duration of exercise (Heck et al., 2003). Whilst the alactic time represents a fictitious period, it is deemed necessary to avoid underestimating the vLamax (Nitzsche et al., 2018b). The time duration to achieve peak power is thought to be equal with the maximal alactic power (Heck et al., 2003).

$$vLa_{max} = \frac{(BLa_{maxpost} - BLa_{pre})}{(t_{test} - t_{alac})}$$

### Equation (2)

Maximal lactate accumulation rate formula  $vLa_{max}$  (mmol·L<sup>-1</sup>·s<sup>-1</sup>) (Mader, 1994).  $BLa_{maxpost}$  = maximum post-exercise blood lactate,  $BLa_{pre}$  = blood lactate prior to the start of the test,  $t_{test}$  = test duration,  $t_{alac}$  = alactic time interval, fictitious time-period where no accumulation of blood lactate occurs.

Recent studies concerning the use of capillary blood sampling to estimate the vLamax, have adjusted parameters of Mader's model implemented precise recommendations dependent upon; exercise modality (Quittmann et al., 2020; Haase et al., 2024), specific methods for determining the alactic time contribution (Dunst et al., 2023b; Meixner et al., 2024), test duration (Langley et al., 2024), consideration of the oxidative metabolism (Yang et al., 2023). It is unclear after which specific time interval capillary blood should be sampled, and for what total duration postexercise to reliably determine the maximum lactate attained (Sperlich et al., 2010; Nitzsche et al., 2018b; Quittmann et al., 2020; Dunst et al., 2023; Langley et al., 2024). Thus, this remains to be clarified. To allow accurate comparisons of results between studies a consensus regarding the most accurate, reliable, and valid measure of vLa<sub>max</sub> specific to each sporting modality need to be considered.

In addition to discrepancies in vLamax test protocols employed, currently, there is no agreed upon denotation of the maximal lactate accumulation rate measured via blood To-date wide sampling. а array of abbreviations, each with distinct connotations to the process involved have been reported including: dLa/dt max (Mader, 1984; Mader & Heck, 1986), VLa'max (Mader, 1994), Vla,max (Mader, 2003), VLamax (Hauser et al., 2014), vLamax (Nitzsche et al., 2020; Meixner et al., 2024), ċLamax (Quittmann et al., 2022), ċLamax (Dunst et al., 2023a; 2023b), and vLa.max (Pohl et al., 2024). A common census is needed when discussing maximal lactate accumulation rate, to build cohesive research environment.

The primary aim of this systematic review is to evaluate the current methodological approaches applied to test an athletes vLamax within different exercise modalities. Secondary we aim to assess the reliability of the vLamax including the alactic time component, and the effects of training interventions. Subject to the review findings we aim to make practical recommendations for future researchers and applied practitioners. These recommendations should provide standardisation procedures allowing for accurate comparisons to be drawn, whilst aiming to enhance the reliability and validity of vLamax assessment. Lastly, we aim to provide a sound rationale for standardisation of an appropriate abbreviation discussing when maximal lactate accumulation rate.

### 2 Material and Methods

### 2.1 Search Strategy

This systematic review was conducted in accordance with the PRISMA (Preferred Reporting Items, for Systematic Reviews and meta - analyses) guidelines (Moher et al., 2009). A systematic literature search using the following databases, PubMed, SCOPUS, Google Scholar, and SPOLIT, was undertaken. As mentioned above Mader (1984) is widely recognised as the inception of the vLamax formula, thus 1984 was used as the earliest available article. The search was conducted between 1984 until July 2024. All database searches were conducted between the 27th - 28th July 2024. The key search terms used were a combination of "vLamax"; "maximal lactate"; "maximal lactate accumulation"; "maximal lactate accumulation rate"; "maximal lactate formation"; "maximal lactate production"; "maximal glycolytic rate"; "maximal glycolytic flux"; "all out exercise" AND "lactate"; "maximal exercise" AND "lactate accumulation"; "maximal lactate" "sprint"; "maximal lactate" AND "cycling"; "maximal lactate" AND "running"; "maximal lactate" AND "swimming"; "maximal lactate" AND "rowing", and the German equivalence.

Therefore, a total of 14 separate searches were conducted across each of the databases. In addition, a reverse search of all papers which have citied Mader's (1984) seminal paper were undertaken using Google Scholar's 'cited by' tool. Furthermore, the same reverse strategy was applied for Heck et al. (2003) paper from which the vLamax formula is frequently cited. All titles and abstracts were examined and screened for eligibility. Screening the reference lists of eligible studies identified an additional four articles. Articles were searched for studies on humans, free full text in either English or German, and published between our date range of 1984 – July 2024.

Articles were pooled together from the four databases and screened as one succinct list. Literature search, identification, and review was conducted by all four authors (JL, NN, RH, MP). Articles were screened at every stage by at least two of this manuscript's authors for eligibility, following the procedure outlined in Figure 1. Any disagreements between authors were resolved by the consensus of the other authors.

#### 2.2 Inclusion and Exclusion criteria

Pre-determined inclusion and exclusion criteria were followed during screening. 1) Original studies using the vLamax formula, 2) written in the English or German language, and 3) exercise duration less than 60 s were included. Whereas original articles that used or mentioned the following were excluded; 1) populations, non-human 2) populations, 3) not in vitro, and 4) mentions of other lactate protocols (maximum lactate steady state), 5) articles that have not been peer reviewed or not original data (systematic review, meta-analysis, conference abstract, postgraduate thesis, book) 6) articles that were primarily mathematical simulations rather than human participants.

#### 2.3 Data Extraction and Analysis

Full text of all included articles were read and the study design, sample size, test duration, exercise modality, vLa<sub>max</sub> formula, methodological considerations, and main conclusion, were extracted and summarised in Tables 1 - 3. There was no blinding to study author, institution, or journal at this stage. Data extraction was carried out by JL, RH, and MP, where NN checked the extracted data for accuracy and totality.

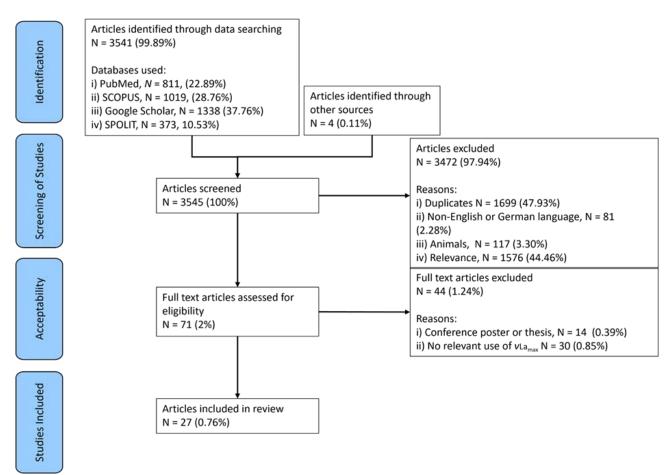


Figure 1. Study selection criteria for systematic review.

### 3 Results

Our comprehensive literature search and screening process identified twenty-seven articles that described the use of the  $\nu$ Lamax formula within their methods, outlined in Table 1. The four main outcomes for screening the articles were: what alactic method within the  $\nu$ Lamax formula was used, exercise duration, exercise modality, and specific methodological considerations. Additionally, studies that assessed the  $\nu$ Lamax reliability (Table 2), and training intervention studies (Table 3) were included.

The literature collectively uses the Mader's (1994)  $\nu La_{max}$  formula, whilst the alactic

component of the equation is commonly modified. In total nine different methods to determine the alactic time span were identified across 27 studies including;

- time to peak power (N = 6),
- time to decline of 3.5% in peak power output (N = 11),
- exercise test duration (s)  $\cdot 0.0909 + 2.0455$  (N = 4),
- fixed time intervals of 3- (N = 2), 3.5- (N = 3), 4- (N = 2), and 5 s (N = 1) after the onset of the sprint,
- time to peak plus oxidative component (N = 1) and the time of fatigue-free force velocity profile (N = 2).

**Table 1.** Articles that have tested an athletes  $\nu$ Lamax.

Identifier	Sample Size and Gender	Training Status	Modality	Test Duration/ distance	Alactic time period	νLa <sub>max</sub> data	BLC Sampling	Summary
Hauser et al. 2014	N = 13 (M13)	Amateur	Cycling	15s	TTP-3.5%	$0.91 \pm 0.18$	0-9 mins every 1 min	
Adam et al. 2015	N = 23 (6F M17)	Amateur	Cycling	15s	TTP-3.5%	$0.72 \pm 0.13$ (T1) $0.72 \pm 0.14$ (T2) $0.70 \pm 0.14$ (T3)	Every 1 min for 10 mins	
Manunzio et al. 2016	N = 4 (M4)	Well Trained	Cycling	15s	TTP	0.54 ± 0.16 (120rpm T1) 0.56 ± 0.16 (120rpm T2) 0.46 ± 0.11 (120rpm T3)	Every 1min for 10 mins	
Yang et al. 2023	N = 10 (M10)	Elite Track	Cycling	15s	TTP-3.5% TTP TTP+tOxi	$0.97 \pm 0.18$ $0.85 \pm 0.12$ $0.88 \pm 0.13$	0-10 mins every 1 min	Only one to use oxid
Dunst et al. 2023a	N = 12 (F3 M9)	Elite Track	Cycling	3s 8s 12s	tFf	0.70 0.76 0.78	0, 1, 3, 5, 7, 10, 15, 20, 25, 30 mins	
Dunst et al. 2023b	N = 9 (M9)	Elite Track	Cycling	3s 8s 12s 60s	tFf	0.70 0.80 0.82 0.31	0, 1, 3, 5, 7, 10, 15, 20, 25, 30 mins	
Harnish et al. 2023	N = 20 (F12 M18)	Amateur	Cycling	15s	TTP-3.5%	0.67 ± 0.02 (T1) 0.64 ± 0.24 (T2)	Every 2 mins from 60s until peak minus 1 mmol · L <sup>-1</sup>	
Poffé et al. 2024	N = 31 (F12 M19)	Recreational (17), Amateur (10) to Elite (4)	Cycling	15s	TTP-3.5%	0.56 ± 0.15 (130rpm)	Every 1min for 10 mins	
					TTP-3.5s	$0.55 \pm 0.14$ (T1) $0.54 \pm 0.13$ (T2)		
Meixner et al. 2024	N = 50 (F20 M30)	Amateur	Cycling	15s	TTP	$0.50 \pm 0.11 \text{ (T1)}$ $0.49 \pm 0.13 \text{ (T2)}$	Every 1min for 10 mins	
					TTP-3.5%	$0.56 \pm 0.13$ (T1) $0.53 \pm 0.14$ (T2)		
Haase et al. 2024	N = 13 (M13)	Amateur	Cycling	10s	3s	0.63 ± 0.14 (T1 - 90rpm) 0.76 ± 0.13 (T2 - 110rpm) 0.86 ± 0.16 (T3 - 130rpm) 0.88 ± 0.15 (T4 - 150rpm) 0.94 ± 0.14 (T5 - 170rpm)	30 s interval until 9 mins, 1 min intervals until 15 mins and 17, 19, 21, 24, 27, 30 mins	

	N = 15			10s		$0.86 \pm 0.17 (10s)$		
Langley et al. 2024	(M15)	Well Trained	Cycling	15s	TTP-3.5%	$0.68 \pm 0.18  (15s)$	Every 1min for 10 mins	
	(1113)			30s		$0.45 \pm 0.07 (30s)$		
						$0.60 \pm 0.16$ (T1 CYC)		
Quittmann et al.	N = 18	Amateur	Cycling /	15s	TTP-3.5%	$0.60 \pm 0.15$ (T2 CYC)	Every 1 min for 10 mins	
2021b	(3F 15M)	Triathlete	Running	138	111-3.3%	0.72 ± 0.16 (T1 100m run)	Every 1 mm for 10 mms	
						0.71 ± 0.16 (T2 100m run)		
	NI 14			CY - 15s	TTD 0 50/ (C)/)	0.01 . 0.00	20 :	
Nitzsche et al. 2018a	N = 14	Amateur	Cycling /	IFL - $16.1 \pm 2.0$ s	TTP-3.5% (CY)	$0.81 \pm 0.09$	30 s interval until 3 mins, 1 min	
	(M14)		IFL	(10 reps)	TTP-3.5% (IFL)	$0.28 \pm 0.09$	interval until 9 mins	
						0.53 ± 0.14 (T1 CYC)		
Quittmann et al.	N = 18	National	Cycling/ Handcycling	15s	TTP	$0.52 \pm 0.14$ (T2 CYC)	Every 1min for 10 mins	
2021a	(F3 M15)	Triathletes				$0.31 \pm 0.09 (T1 HCY)$		
						$0.32 \pm 0.10$ (T2 HCY)		
N = 2	N = 25	A 1	ъ :	15.		$0.47 \pm 0.06$ (Amateur)	0 1 1	Predicting
Hanon et al. 2011	(F6 M19)	Amateur	Running	15s		$0.59 \pm 0.11$ (Elite)	0- & 4-mins post	νLa <sub>max</sub>
		J	Running	8-14s	TTP	0.65 ± 0.23 (8s Tk T1a)	0-10 mins every 1min	
						$0.64 \pm 0.22$ (8s Tk T1b)		
						$0.61 \pm 0.23  (10s  Tk  T1a)$		
						0.56 ± 0.19 (10s Tk T1b)		
						$0.60 \pm 0.22$ (12s TK T1a)		
						$0.60 \pm 0.21$ (12s TK T1b)		
						$0.60 \pm 0.19$ (14s TK T1a)		
Wawer et al. 2020						$0.59 \pm 0.19$ (14s TK T1b)		
						$0.84 \pm 0.15$ (10s Trm T 2a)		
						$0.83 \pm 0.22$ (10s Trm T2b)		
						$0.91 \pm 0.17$ (12s Trm T2a)		
						$0.91 \pm 0.18$ (12s Trm T2b)		
						$0.74 \pm 0.21  (10s  TK  T3a)$		
						$0.71 \pm 0.20 \text{ (10s Trm T3b)}$		
0	NT			13.90 ± 1.42s	·E 0.0000			
Quittmann et al.	N = 16	Competitive	Running	$13.89 \pm 1.47$ s	tExer · 0.0909 +	0.79 ±0.18 (100m)	Every 1 min for 10 mins	
2020	(F5 M11)	MD	Ü	$13.86 \pm 1.47$ s	2.0455	,	,	
0 : 1	N = 44				tExer · 0.0909 +	$0.74 \pm 0.14$ (M)		
Quittmann et al.	11 - 44		Running		CLACI O.O.O.	0.7 1 = 0.11 (141)	Every 1 min for 10 mins	

Thron et al. 2024	N = 34 (F15 M19)	Recreational	Running	100m	TTP	$0.92 \pm 0.20$ (100m athletes) $0.83 \pm 0.16$ (400m athletes) $0.71 \pm 0.13$ (800m athletes)	Every 1 min for 10 mins
Pohl et al. 2024	N = 21 (F8 M13)		Running		tExer · 0.0909 + 2.0455	0.59 ± 0.09 (Baseline) 0.51 ± 0.01 (High BLC) 0.53 ± 0.10 (CHO Low) 0.54 ± 0.10 (CHO High) 0.57 ± 0.10 (CHO Acute)	Every 1 min for 10 mins
Schünemann et al. 2023	N = 10 (F3 M7)	National level	Rowing	10s	tExer · 0.0909 + 2.0455	$0.45 \pm 0.14$	0-10 mins every 1 min
Held et al. 2024	N = 17 (F8 M9)	Amateur	Rowing	20s	4s	$0.29 \pm 0.11 \text{ (T1)}$ $0.28 \pm 0.10 \text{ (T2)}$	Every 1 min for 10 mins
Nitzsche et al. 2018b	N = 32 (M32)	University Students	Isokinetic Force Loads	8 reps T1: $12.1 \pm 1.02$ s T2: $11.5 \pm 1.03$ s 16 reps T1: $23.1 \pm 0.31$ s T2: $23.0 \pm 0.28$ s	TTP-3.5%	0.25 ± 0.11 (T1 8reps) 0.27 ± 0.11 (T2 8reps) 0.27 ± 0.07 (T1 16reps) 0.26 ± 0.07 (T2 16reps)	30s interval until 3 mins, 1 min interval until 9 mins
Nitzsche et al. 2020	N = 24 (M24)	University Students	Isokinetic Force Loads	10 reps (15s)	3s	$0.27 \pm 0.07$ (T1 HVLL) $0.30 \pm 0.07$ (T2 HVLL) $0.25 \pm 0.12$ (T1 LVHL) $0.29 \pm 0.09$ (T2 LVHL)	30s interval until 3mins, 1 min interval until 9 mins
Zwingmann et al. 2020	N = 8 (M8)	National	Canoe polo	15s	3.5s	$0.58 \pm 0.10$	Every 1 min for 10 mins
Quittmann et al. 2018	N = 12 (M12)	National Triathletes	Handcycling	15s	TTP-3.5%	$0.45 \pm 0.11$	Every 1 min for 10 mins
Mavroudi et al. 2023	N =14 (F6 M8)	Semi Elite	Swimming	25m: 11.75 ± 1.38 35m: 17.76 ± 2.04 50m: 26.78 ± 3.21	3.5s 4s 5s	$0.75 \pm 0.18$ $0.54 \pm 0.18$ $0.49 \pm 0.16$	every minute until peak

Legend: RCT; randomised control trial, CS; case study, T; Trial, IFL; Isokinetic Force Load, CYC; Cycling, CHO; Carbohydrate, RPM; Revolutions per minute, min; minute, s; Seconds, M; Males, F; Females, HVLL; high volume low load, LVHL; low volume high load, tFf; time fatigue free force, tExer, time to finish the test/exercise, TTP; Time to Peak Power, TTP-3.5%; time until peak power dropped by 3.5 %, TK; track, Trm; Treadmill, MD; Middle distance, HCY; Handcycling, BLC; Blood lactate concentration, N; Sample size, %; Percentage, vLamax; maximal lactate accumulation rate.

**Table 2.** Articles investigating the vLa<sub>max</sub> reliability and/or variability.

Identifier	Sample Size and Gender	_	Modality	Test Duration/ distance	Alactic time- period	Reliability/Variability results -	· Reliability/Variability results – νLa <sub>max</sub>	Reliability/Variability results – BLC
Adam et al. 2015	N = 23 (6F M17)	Amateur	Cycling	15s	TTP-3.5%	ICC = 0.881; RMSE= 0.25; CV = 5.8%	ICC = 0.904; RMSE = 0.045; CV = 6.3%	BLC <sub>pre</sub> : ICC = 0.804; RMSE = 0.20 mmol · L-¹; CV = 18.8 % BLC <sub>peak</sub> : ICC = 0.856; RMSE = 0.58; CV = 6.8 % ΔBLC: ICC = 0.891; RMSE = 0.52; CV = 7.0 %
Harnish et al. 2023	N = 20 (F12 M18)	Amateur	Cycling	15s	TTP-3.5%	CV = 38.3%	ICC = 0.66; CV = 18.6%	BLC <sub>pre</sub> : CV = 45.6 % BLC <sub>peak</sub> : CV = 23.3 %
					3.5s	3.5s: fixed	3.5s: ICC = 0.91; SEM = 0.02; CV = 3.1%	
Meixner et al. 2024	N = 50 (F20 M30)	Amateur	Cycling	15s	TTP	TTP: ICC = 0.41; SEM = 0.71; CV = 30%	TTP: ICC = 0.87; SEM = 0.06; CV = 12.1%	ΔBLC: ICC = 0.911; SEM = 0.22; CV = 3.6 %
					TTP-3.5%	TTP-3.5%: ICC = 0.52; SEM = 1.83; CV = 51%	TTP-3.5%: ICC = 0.79; SEM = 0.15; CV = 26.4%	
Quittmann et al. 2021b	N = 18 (3F 15M)	Amateur Triathlete	Cycling / Running	15s	TTP-3.5%	CYC: ICC = 0.592 100 m run: ICC = 0.242	CYC: ICC = 0.894; Bias (LoA) = -0.003 (-0.149 to 0.143) 100 m run: ICC = 0.868; Bias (LoA) = -0.022 (-0.181 to 0.137)	CYC: BLC <sub>pre</sub> : ICC = 0.537 BLC <sub>peak</sub> : ICC = 0.870 ΔBLC: ICC = 0.867 100m run: BLC <sub>pre</sub> : ICC = 0.335 BLC <sub>peak</sub> : ICC = 0.808 ΔBLC: ICC = 0.783
Quittmann et al. 2021a	N =18 (F3 M15)	National Triathletes	Cycling/ Handcycling	15s	ТТР	CYC: ICC = 0.525 HCY: ICC = -0.115	CYC: ICC = 0.872; LoA = -0.15 to 0.14 HCY: ICC = 0.828; LoA = -0.10 to 0.12	able. ree (iii do
Nitzsche et al. 2018b	N = 32 (M32)	University Students	Isokinetic Force Loads	8 reps T1: 12.1 ± 1.02s T2: 11.5 ± 1.03s 16 reps T1: 23.1 ± 0.31s T2: 23.0 ± 0.28s	TTP-3.5%	8 reps: r = 0.67; Bias (LoA) = - 0.02 (Median) (-0.33 to 1.96) s 16 reps: r = 0.48; Bias (LoA) = 0.11 (-6.59 to 6.81) s	8 reps: r = 0.72; Bias (LoA) = 0.02 (-0.09 to 0.13) 16 reps: r = 0.68; Bias (LoA) = - 0.008 (-0.118 to 0.102)	BLC <sub>peak</sub> : 8 reps: r = 0.688; 16 reps: r = 0.821

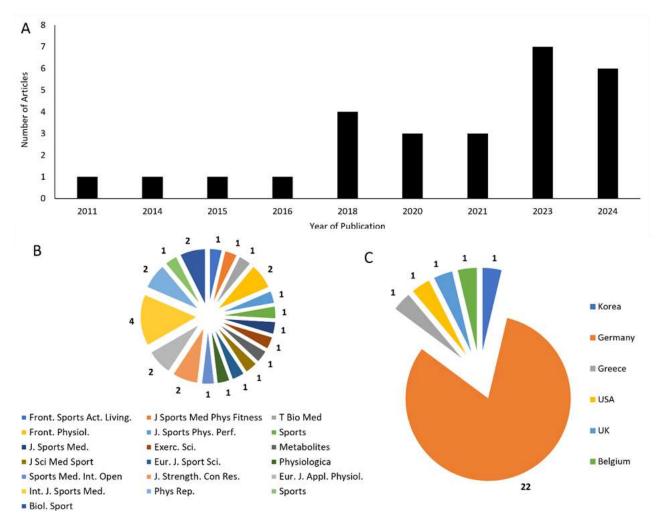
Held et al. 2024	N = 17 (F8 M9)	Amateur	Rowing	20s	4s	- ICC = 0.85; SEM = 0.02	BLC <sub>peak</sub> : ICC = 0.88; SEM = 0.3
Wawer et al. 2020	N = 73 (F12 M61)	University Students	Running	8-14s	TTP	8s Tk: ICC = 0.73, CV = 11.1% 8s Tk: ICC = 0.89, CV = 9.8% 10s Tk: ICC = 0.16, CV = 10.1% 10s Tk: ICC = 0.82, CV = 12.9% 12s TK: ICC = 0.76, CV = 8.7% 12s TK: ICC = 0.92, CV = 9.0% 14s TK: ICC = 0.11, CV = 10% 14s TK: ICC = 0.84, CV = 10.7% 10s Trm: ICC = 0.47, CV = 10.2% 10s Trm: ICC = 0.76, CV = 7.6% 12s Trm: ICC = 0.26, CV = 7.8% 12s Trm: ICC = 0.79, CV = 6.1%	10s Trm: BLC <sub>peak</sub> : ICC = 0.91; CV = 5.2 %  12s Trm: BLC <sub>peak</sub> : ICC = 0.77; CV = 5.8 %

Legend: BLC<sub>pre</sub>; blood lactate concentration pre-test, BLC<sub>peak</sub>; peak blood lactate concentration post-test,  $\Delta$ BLC; change of blood lactate concentration (BLC<sub>peak</sub>-BLC<sub>pre</sub>), CV; coefficient of variation, CYC; Cycling, F; Female, HCY, Handcycling, ICC; Intraclass correlation coefficient, LoA; Limits of Agreement, M; Male, min; minute, N; Sample size, r; Pearson's r correlation coefficient, reps; repetitions, RMSE; Root Mean Square Error, s; seconds, SEM; Standard Error of Measurement, T; Trial, TK; track, Trm; Treadmill, TTP; Time to Peak, TTP-3.5%; time until peak power dropped by 3.5 %,  $\nu$ La<sub>max</sub>; maximal lactate accumulation rate, %; Percentage.

**Table 3.** Articles that have measured the adaptation of  $\nu$ Lamax following a training intervention.

Identifier	Sample Size and Gender	Training Status	Modality	Test Duration/ distance	Alactic time-period	Training Intervention	Sessions per week	Duration (weeks)	νLa <sub>max</sub>	ΔυLa <sub>max</sub>
Hommel et al. 2019	N = 30 9 (M, 10 per group)	amateur cyclists	Cycling	15s	TTP-3.5%	SIT: not standardized ET: not standardized	3	6	Pre – SIT: 0.76 (0.18) ET: 0.75 (0.18) Post – SIT: 0.63 (0.15) ET: 0.74 (0.17)	SIT: -10.53%, sig. reduction (-0.08 (p < 0.05)) ET: -1.35%, no sig. changes
Nitzsche et al. 2020	N = 24 (M, HVLL = 14; LVHL = 10)	healthy strength trained	Isokinetic Force Loads	10 reps (15s)	3 s	5 sets per exercise, 90 s break HVLL: as much repetitions as possible at 50% 1RM LVHL: 10 repetitions at 70% 1RM Exercises: leg press, leg extension, leg flexor (prone position)		6	Pre – HVLL: 0.271 (0.067) LVHL: 0.249 (0.122) Post – HVLL: 0.298 (0.067) LVHL: 0.291 (0.089)	HVLL: +9.96% (p = 0.022, d = 0.406), LVHL: +16.87% (p = 0.233, d = 0.384)

Legend: ET; endurance training, HVLL; high volume low load, LVHL; low volume high load, M; Male, N; Sample size, 1RM; one repetition maximum, reps; repetitions, s; seconds, SIT; sprint interval training, TTP-3.5%; time until peak power dropped by 3.5 %, vLa<sub>max</sub>; maximal lactate accumulation rate, %; Percentage.



**Figure 2.** Summary of the article extracted for this review (N = 27). A- Distribution of articles via year the articles were published, B- Distribution of articles via journal of publication, C- Distribution of articles via first author country of origin.

From the included articles vLamax was tested across several different populations of varying performance levels, ages, sex, and across an array of modalities. The included articles spanned from 2011 to and including 2024 (Figure 2A). The articles were published in a variety of journals with no conceivable pattern (Figure 2B). The articles were authored primarily by German based researchers (22 in total), with the remaining from Belgium, Korea, Greece, UK, and the USA (Figure 2C; (Hauser et al., 2014, Adam et al., 2015, Manunzio et al., 2016, Yang et al., 2023, Dunst et al., 2023a, Dunst et al., 2023b, Harnish et al., 2023, Poffé et al., 2024, Meixner et al., 2024, Haase et al., 2024, Langley et al., 2024, Quittmann et al., 2021b, Nitzsche et al., 2018a, Quittmann et al., 2021a, Hanon et al., 2011,

Wawer et al., 2020, Quittmann et al., 2020, Quittmann et al., 2023, Thron et al., 2024, Pohl et al., 2024, Schünemann et al., 2023, Held et al., 2024, Nitzsche et al., 2018b, Nitzsche et al., 2020, Zwingmann et al., 2020, Quittmann et al., 2018b, Mavroudi et al., 2023). A total sample size outlined was N = 580, with (N = 452)coming from amateur, well-trained (N = 63), and elite (N = 65) participants (as outlined by journal reported demographics), whereby 129 of the 580 participants were female (22.24%) and the other 451 participants were male (77.76%). The average age of the participants was 23 years of age, with two studies using populations under the age of 18 years old (11-15-year-olds). Seven different modalities were tested – cycling (N = 14), swimming (N = 1), running (N = 7), handcycling (N = 2), kayaking (N = 1), rowing (N = 2), and Isokinetic force load (N = 3). Studies were included twice if they tested multiple modalities. Exercise duration ranged from 3 to 60 seconds, with the most popular test duration of 15 s (N = 15). Additionally, a variety of study designs were outlined, 17 studies were classified as cohort studies, 8 papers identified reliability and 2 papers identified the effects of training interventions on  $\nu La_{max}$ .

Time to peak blood lactate concentration (tBLC<sub>max</sub>) is a key variable for  $\nu$ La<sub>max</sub> research, yet only 6 of the 27 studies documented their respective tBLC<sub>max</sub>. Within cycling research articles,  $\nu$ La<sub>max</sub> test durations of 10, 15, and 30 s elicit tBLC<sub>max</sub> durations of  $5 \pm 2$  mins,  $4.36 \pm 2.4$  mins, and  $7 \pm 2$  mins, respectively. An average tBLC<sub>max</sub> for Isokinetic force  $\nu$ La<sub>max</sub> tests were  $2.31 \pm 0.7$  mins. Additionally, a 20 s rowing  $\nu$ La<sub>max</sub> test elicited average tBLC<sub>max</sub> values of  $5.15 \pm 0.5$  mins. Lastly, swimming  $\nu$ La<sub>max</sub> tests of 25 m, 35 m, and 50 m elicited tBLC<sub>max</sub> values of  $2.2 \pm 0.8$  mins,  $2.1 \pm 0.7$  mins, and  $2.4 \pm 1.0$  mins, respectively.

### 4 Discussion

This systematic review examined all published research applying the vLa<sub>max</sub> within exercise durations <60 s and in human populations since its inception 40 years ago by Mader (1984). A total of 27 studies including 580 participants were included within the analysis across seven different modalities spanning test durations of 3 - 60 s. Additionally, this review evaluated the alactic time component, test duration, modality, reliability, training interventions, blood lactate sampling period, on the vLa<sub>max</sub> and proposed future test recommendations.

Over the last 40 years, the literature discussing the maximal lactate accumulation rate has used a wide variety of abbreviations, each having distinct connotations to the

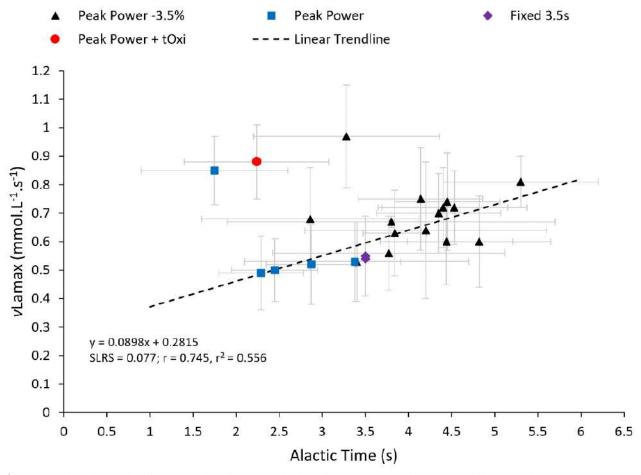
process in question. The most common terminologies used when discussing maximal lactate accumulation rate are; dLa/dt max (Mader, 1984; Mader & Heck, 1986), VLa'max (Mader, 1994), vLa,max (Mader, 2003), VLamax (Hauser et al., 2014), vLamax (Nitzsche et al., 2020; Meixner et al., 2024), ċLamax (Quittmann et al., 2022), vLamax (Dunst et al., 2023a; 2023b), vLa.max (Pohl et al., 2024), and recently vLapeak (Wackerhage et al., 2025). The rate of glycolysis is activated as a function of free ADP and AMP, which in turn regulates the rate of PFK. The ' $\nu$ ' denotes the rate of the reaction in with the Michaelis-Menten accordance Kinetics (Michaelis & Menten, 1913; Cornish-Bowden, 2014). It should be noted that the use of the 'v' does not imply that the actual reaction rate is measured during the all-out sprint tests. The term Lamax/Lapeak denotes the highest lactate accumulation in the blood post exercise. The term 'vLapeak' has been suggested due to the unlikely occurrence PFK activity and thus glycolysis is maximally activated in vivo due to decrease in pH during all out exercise inhibiting the PFK activity (Dobson et al., 1986) and ADP/AMP concentrations being unlikely to rise high enough to maximally elicit PFK activity (Wackerhage al., 2025). Additionally, the peak rate of glycolysis and lactate synthesis only occurs for a short timeperiod prior to declining, therefore the maximal rate of glycolysis is not sustained over the time-course of the exercise (Mader et al., 2002; Heck et al., 2003; Porter & Langley, 2025). A common census is needed when discussing maximal lactate accumulation rate, to build cohesive research environment.

### 4.1 Alactic time

The alactic time span is considered to reflect the "lactate-free period" where accumulation of lactate in the blood is negligible (Mader, 1994). Whilst a timespan where no glycolytic activity occurs is fictitious (Brooks, 2018; Chung et al., 1998), the alactic period represents a crucial element of the vLamax calculation with the exclusion leading to underestimating the vLamax (Nitzsche et al., 2018b). As the alactic time is used to determine the denominator in the vLamax equation variations significantly influence the vLamax, with a variation of 1 second influencing the vLamax by up to 26% when assessed via a 15 s sprint (Hauser, 2014). Therefore, a valid and reliable method to identify the alactic time is essential.

Currently, nine methods to determine the alactic period have been applied with varying approaches both within and between modalities (Table 1). Figure 3 highlights the

influence of the alactic method on the vLamax over a 15 s cycling test. Unsurprisingly, a strong correlation between vLamax and alactic durations were observed (Figure 3). On average, TTP-3.5% produced the longest alactic time  $(4.16 \pm 0.61 \text{ s})$ , compared with a fixed time of 3.5 s, and TTP (2.75  $\pm$  0.49 s). Yang et al. (2023) results were excluded from the group linear analysis, due to higher peak vLamax values likely attributed by the elite population (national level sprint cyclists). Yang et al. (2023) results observe the same trend in vLamax and alactic duration. The longest alactic time was associated with TTP-3.5% (3.28  $\pm$  1.08 s) compared with TTP + oxidative time (2.24 ± 0.84 s), and TTP ( $1.75 \pm 0.59 \text{ s}$ ) (Figure 3).



**Figure 3.** The relationship between the alactic method and  $\nu$ La<sub>max</sub> assessed via 15 s all-out cycle ergometry testing. Individual markers represent the mean  $\nu$ La<sub>max</sub> reported within each study (n = 20). Vertical error bars denote  $\nu$ La<sub>max</sub> standard deviation. Horizontal error bars denote alactic time standard deviation. \* *Note Yang et al.* 2023 (n = 3) results were excluded from the linear regression.

The alactic time-period is thought to be identical to the TTP, with researchers aiming to identify the corresponding period (Heck et al., 2003). Subsequently, the alactic method 'TTP-3.5%' was introduced to account for a 2.5% measurement error of the SRM power meter 2003). Fixed duration timespans are derived from Heck et al's. (2003) computer simulation models. Heck et al. (2003) reports that the alactic interval to be dependent upon test duration, with the alactic period of 3, 4, and 8 s corresponding to test durations of 10.5, 21.5, and 45 s, respectively. Dependent on the method of determination, modality, and participant population the reliability of the alactic time-period assessed via intraclass correlation coefficient (ICC) ranged from good (0.881) to poor (0.115) (Table 2). Within cycling the reliability of the TTP-3.5% was moderate to good (TTP-3.5% ICC: 0.52 - 0.881), and only poor to moderate TTP (ICC: 0.41 – 0.525) (Table 2). Applying a fixed alactic time overcomes the reliability challenges. A fixed time interval has been demonstrated to enhance the ICC (0.91) of the vLamax results compared with alactic methods; TTP (ICC = 0.87), and tPP-3.5% (ICC = 0.79) (Meixner et al., 2024).

However, there are distinct limitations of applying a fixed time interval dependent on the test duration. 1) To accurately determine the maximal glycolytic flux of an athlete, exercise should be all-out in nature with no pacing employed. Therefore, strategies maximal exercise independent of test duration should provoke the same alactic timespan. Moreover, if an athlete is employing pacing strategies the test duration is too long. 2) A fixed alactic timespan is not reflective of the individual's physiology. An individual's muscle fibre typology may influence the alactic time-duration with athletes who possess a higher percentage of type II fibres able to metabolise more PCr and yield ATP more rapidly than athletes with high percentages of type I fibres (Esbjornsson-Liljedahl et al., 1999; Greenhaff et al., 1994).

As the alactic time-period reflects a fictitious timespan where no glycolytic activity occurs (Brooks, 2018; Chung et al., 1998), alternative approach of negating the alactic timespan from the vLamax equation could be employed. Such an approach would enhance the reliability of the vLamax in line with applying a fixed time-period as reported by Meixner et al., (2024). However, it should be considered that removing the alactic timespan would reduce the calculated vLamax as the denominator in the equation would be increased. Subsequently, removing the alactic timespan may underestimate the maximal glycolytic rate (Nitzsche et al., 2018b) and subsequently increase the calculated maximal lactate steady state when employing Mader's model of human energy metabolism (Mader, 1984; Mader & Heck, 1986; Mader, 2003; Hauser et al., 2014; Wackerhage et al., 2023; Poffé et al., 2024).

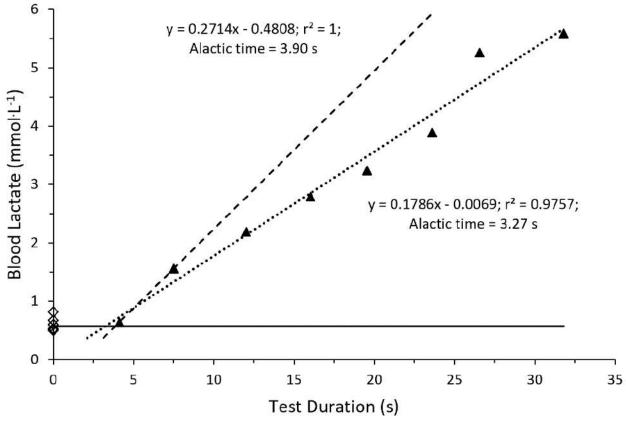
Recently, Dunst et al. (2023) has proposed an attractive alternative for measuring the alactic period within cycling derived from the individual's fatigue-free force velocity profile. In cycling the power output is reflected by the parabolic relationship between mechanical force and pedaling rate. Subsequently, if the initial inertia resistance is set too low the time to achieve peak power will be reduced underestimating the alactic timespan. Contrary, if the inertia resistance is too high the time to achieve peak power will be extended and the alactic timespan overestimated (Dunst et al. 2023a). Dunst et al. (2023a) proposes the time point of the first systematic deviation from the fatigue-free force velocity profile should identify the alactic timespan by reflecting the end of maximal energy flux and should correspond with a non-significant accumulation of blood lactate. A valid forcevelocity profile can be derived from testing at the two extreme ends of the F/v profile (Sašek et al., 2022). Dunst et al. (2023a) utilised a motoric 6 s sprint against the lowest cycling resistance possible to attain maximal cadence and the initial 3-4 pedal revolutions during the all-out sprint to generate the fatigue-free forcevelocity and power-velocity profiles. The time of fatigue free force (tFf) was defined as, 'the time where the force-velocity profile decreased below the fatigue-free maximum and never returned'. Extrapolating the linear regression of the  $\Delta BLC$  from the 3, 8, and 12 s sprints, to a time point where no  $\Delta BLC$  occurs correlated closely with tFf and used to validate this approach.

Whilst Dunst et al. (2023a) tFf provides a potential individualised measure of the alactic timespan in cycling, this approach cannot easily be applied to other modalities where force and velocity cannot be accurately and reliably sampled at a high frequency. Whereas Dunst et al's. (2023a) approach to validate the model may provide a simple and effective method to identify the alactic timespan applicable across modalities. Measurement of ΔBLC following two or more maximal all-out efforts of ≤12 s, and the extrapolation of a simple linear regression to the time point of no accumulation of blood lactate may be used to identify the alactic timespan (Dunst et al. 2023a).

The regression analysis method used by Nitzsche et al. (2017) to determine talac during isokinetic force loads also appears to be helpful. The procedure is based on several tests with different load times to determine the

respective BLC<sub>peak</sub> and the pre-load blood lactate concentration associated with the test; after plotting the BLC<sub>peak</sub> against the test time, a regression line is used to determine the intersection with the regression line of the pre-load blood lactate concentrations (BLC) (Figure 4). The intersection point then indicates the end of the fictitious lactate-free interval and the beginning of increased lactate accumulations. This procedure is based on a method from Danforth (1965), in which the kinetics of PCr and muscle lactate were calculated (Mader 2015).

Presently, no sound recommendations of the 'gold standard' method in identifying the alactic time can be provided. Dunst et al. (2023a) tFf model has only been validated for a small population of elite track sprint cyclists who demonstrated linear F/v profiles. This approach needs to be validated across wider population pools, and the potential application across modalities investigated. Additionally, the reliability of this new approach has yet to be verified and requires further research. Likewise, further studies investigating the reliability and validity of the application of linear regression from maximal BLC are required. In the absence of a validated alactic method of determination we advocate for the adoption of a fixed alactic timespan to enhance the reliability of the vLamax. In accordance with Dunst et al. (2023b) and Nitzsche et al. (2017) experimental findings, Heck et al. (2003) computer simulations and Quittmann et al. (2020) interpolation identify an timespan of ~3 s to be appropriate for a 10 s test duration.



**Figure 4.** Linear regression analysis of blood lactate concentrations (black triangles) to identify the alactic time-period, sample data from a male test subject adapted from (Nitzsche et al., 2017). Maximal BLC measured over various maximum isokinetic strength tests with different load times at an angular velocity of 120°s<sup>-1</sup>. The solid parallel line to the x-axis represents the mean resting blood lactate values. The short-dashed line represents the regression calculated from all BLC. The intercept of the regression for all blood lactate concentration is at 3.27 s and represents the end of the alactic time interval. The intercept denotes the time where no significant increase in blood lactate concentration occurs prior to this point (Nitzsche et al., 2017). The larger dashed line represents the regression from BLC of the first two test durations (4.13 and 7.52 s) where the intersect occurs at 3.90 s.

### 4.2 The Influence of Oxidative Contribution

For the most accurate determination of an individual's vLamax, the interactions of all three energy systems need to be considered. Recently, Yang et al. (2023) demonstrated the importance of factoring in the oxidative metabolic component within the vLamax equation during 15 s all-out cycling (Equation 3). Whilst Yang et al. (2023) only observed a small oxidative contribution (3.13  $\pm$  1.61%) of the total energy supplied over the 15 s sprint, this significantly (p < 0.0001) increased the vLa<sub>max</sub> (ES d = 0.23). Over a 100 m sprint the oxidative metabolism has been demonstrated to contribute ~10% of the total energy expenditure (Park et al., 2021). Furthermore, the oxidative metabolism has been reported to

range from 16-33% during a 30 s Wingate test (Kavanagh and Jacobs, 1988; Smith & Hill, 1991; O'Brien et al., 1997; Beneke et al., 2002). However, a shorter test duration demonstrates a reduction in the oxidative demand thus limiting the impact on the  $\nu$ Lamax (Langley et al., 2024).

Considering the oxidative contribution may enhance the accuracy of the vLa<sub>max</sub> equation and provide insights into interindividual differences in energy metabolism. Whilst the oxidative energy metabolism contribution to single sprint is small (Yang et al., 2023), events where repeated sprint performance is required, such as Olympic level sprint events enhance the reliance on the oxidative metabolism (Lievens et al., 2020).

Recently, Ferguson et al. (2021a) highlighted the linear relationship between power-outputs associated with high reliance on the oxidative metabolism (2-, 8- 20-min power) and 15 and 30 s sprint power. Subsequently, monitoring changes in the oxidative energy supply may highlight deficiencies and provide a basis for training recommendations to enhance sprint power (Ferguson et al., 2021a; 2021b; 2023). However, this approach requires an expensive metabolic cart and additional metabolic calculations to determine both the oxidative and PCr energy metabolism, making this approach practically challenging during field-based testing.

$$Pure vLa_{max} = \frac{BLa_{maxpost} - BLa_{pre}}{t_{test} - (t_{alac} + t_{oxi})}$$

Equation (3)

'Pure'  $\nu L_{amax}$  (mmol  $\cdot$  L<sup>-1</sup>  $\cdot$  s<sup>-1</sup>),  $t_{oxi}$  denotes the percentage of oxidative energy contribution converted into time in seconds (Yang et al., 2023).

### 4.3 Test Duration

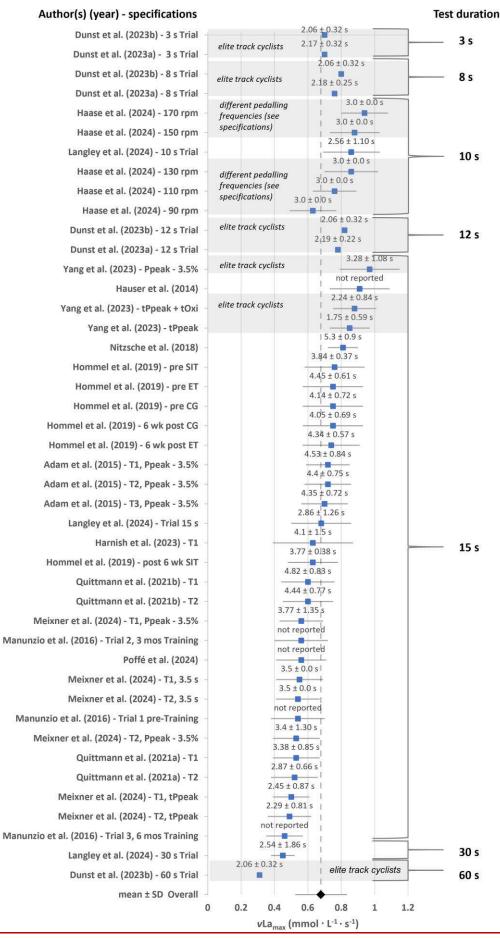
Along with the alactic time-period, the test duration comprises the denominator in the equation and thus significantly  $\nu La_{max}$ influences the results (Mader, 1994). The optimal test duration should be sufficient in length to ensure the maximal rate of glycolytic flux is achieved, however, longer test durations may underestimate vLamax for three key reasons: 1) Glycolysis is self-limiting due to the production of H+ causing a decline in muscle pH, which reduces the metabolic activity of PFK (Erecińska et al., 1995). 2) The longer the test duration the larger the denominator, thus, the smaller the calculated vLamax. 3) Longer test durations may elicit pacing strategies, reducing the F-v and P-v profiles (Robin et al., 2021), and subsequent energy metabolism

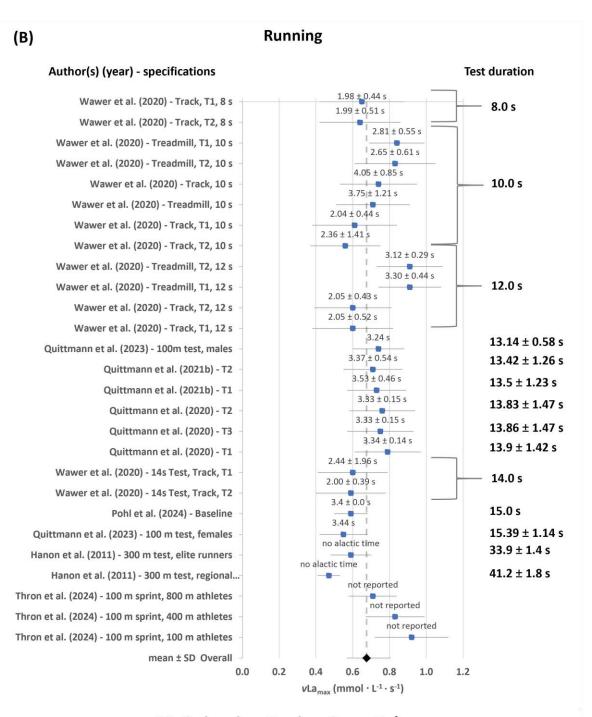
(Hirvonen et al., 1992). However, the shorter the test duration, the larger the influence of the alactic timespan. Where small variations in the alactic duration can lead to large fluctuations in the calculated vLa<sub>max</sub> (Hauser, 2013). This is potentially problematic as this portion of the equation is subject to the largest error, reflected in the variance in the reliability (Table 2 and section 4.5).

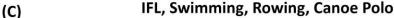
Forest plots illustrated in Figure demonstrate the influence of test duration, alactic time, performance level, and movement velocity on vLamax. The mean vLamax calculated from pooled data dependent on test duration grouped within 1 s identified the highest vLamax was attained between test durations of 10 – 12 s for all modalities (cycling, running, swimming, and rowing) except isokinetic force tests (Figure 6 & 7). Figure 5A highlights both elite cyclists and high movement velocities produce high vLamax values which may confound the results for cycling providing a bias to shorter test durations. Additionally, variations in the alactic times employed between studies makes interpretation of the influence of test duration challenging, as previously highlighted in Figure 3 longer alactic times are associated with a higher νLa<sub>max</sub>.

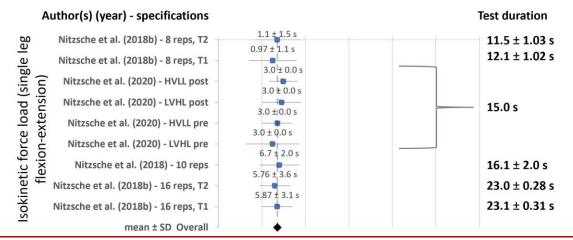
Whilst data is more limited Figure 5C shows the shorter test durations ~10-12 s produced higher vLa<sub>max</sub> values for both swimming and rowing. Contrary, mean vLa<sub>max</sub> values varied by only 0.02 mmol·L<sup>-1</sup>·s<sup>-1</sup> for isokinetic force tests irrespective of test duration (~12 - 23 s). Longer relaxation phases of the knee flexors/extensors and small active muscle mass during isokinetic force testing may explain the small variance in vLa<sub>max</sub> across test durations (Nitzsche et al., 2018b).

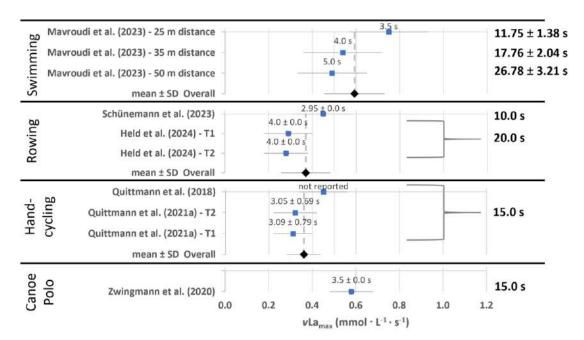
(A) Cycling



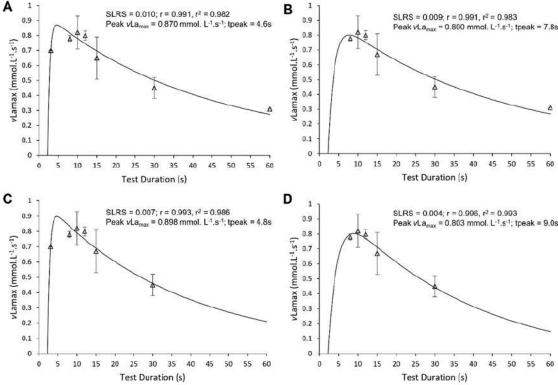




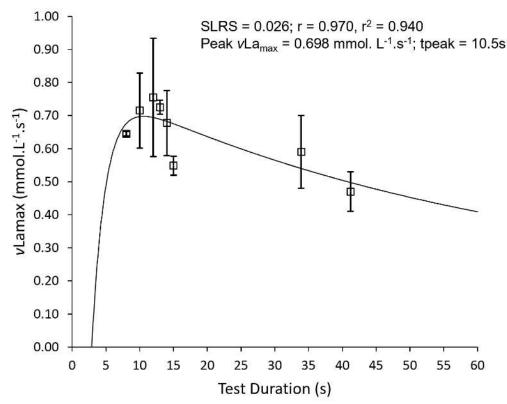




**Figure 5.** Three forest plots illustrate the reported vLa<sub>max</sub> results arranged dependent upon test duration and exercise modality (A) cycling, (B) running, and (C) various modalities including: isokinetic force load (IFL), swimming, rowing, hand-cycling, and canoe polo. Blue squares denote the group mean and standard deviation (SD) highlighted with error bars. Mean and SD alactic times are reported directly above each group vLa<sub>max</sub>. Black diamonds denote the mean values for modality, error bars represent SD. Grey highlights identify possible cofounding factors including performance level and movement velocity.



**Figure 6.** A four-panel plot illustrating data dependent bi-exponential models of cycling  $\nu$ La<sub>max</sub> dependent on the test duration. Triangles denote the mean  $\nu$ La<sub>max</sub> from pooled data, error bars represent the standard deviation, where there was a single study for the test duration raw standard deviation was applied. The alactic time component was constant at 2.12 s for all models in accordance with Dunst et al. (2023a; 2023b). **A)** Bi-exponential model calculated from the sum of least residuals squared (SLRS) using all data points. **B)** Bi-exponential model excluding 3 s test duration  $\nu$ La<sub>max</sub>. **C)** Bi-exponential model excluding both the 3 s and 60 s test duration  $\nu$ La<sub>max</sub>. \*  $\nu$ La<sub>max</sub> from Dunst (2023a) and Dunst (2023b) have been calculated from their mean time of the fatigue-free force-velocity profile representing the alactic time-period, mean ΔBLC, and specific test duration.



**Figure 7.** Running  $\nu$ Lamax bi-exponential model plotted against test duration. A mean alactic time of 2.9s was calculated from the mean values pooled and applied to the bi-exponential model.

The measurement of vLa<sub>max</sub> at specific time points only allows comparisons to be drawn at each specific test duration. Individual data points do not reflect the vLamax across all possible time points. To allow estimations of the vLamax across all possible test durations biexponential models have been constructed using the pooled data for cycling (Figure 6) and running (Figure 7). Bi-exponential modelling predicts a test duration of 10.5 s as optimal to elicit the highest attainable vLamax during running (Figure 7). Four separate biexponential models have been constructed for cycling with the exclusion of the data points for 3 and 60 s (Figure 6). All bi-exponential models demonstrate excellent fit (r<sup>2</sup> ≥ 0.982) and predict the optimal test duration to elicit the highest vLa<sub>max</sub> ranges from 4.6 – 9.0 s (Figure 6). Whilst model D predicting peak vLamax to occur after 9.0 s offers the best fit ( $r^2 = 0.993$ ) this is likely due to the inclusion of fewer data points. Models C and D exclude the 60 s test data point as this is beyond the recommended maximum test duration of 30 s (Heck et al., 2003). This inclusion may elucidate more accurate blood lactate kinetics. Models B and D exclude the 3 s data point where vLamax is constructed from a small mean ΔBLC 0.62 mmol·L<sup>-1</sup>, very short time denominator of 0.88 s and elite cohort (Dunst et al., 2023a; 2023b). Subsequently, minor changes will significantly influence the calculated vLamax. Nevertheless, maximal glycolytic rate may occur earlier than previously predicted for testing vLamax with Dunst et al. (2023a) demonstrating a nearly linear rate of lactate accumulation in the blood between 3, 8, and 12 s sprints. These findings are supported by Hirvonen et al. (1987) who observed a linear ΔBLC during sprint running over distances from 40-100 m.

As the vLa<sub>max</sub> is dependent upon the test duration (Langley et al., 2024), a single test only corresponds to a single time point. With this consideration practitioners and researchers must ensure a standardised test duration to allow reliable comparisons. Furthermore,

without knowing the test duration for the individual which elicits the highest possible  $\nu La_{max}$  a denotation of  $\nu La_{peak}$  maybe more appropriate.

Furthermore, whilst utilising the vLamax and VO<sub>2max</sub> as input parameters within Mader's model of human energy metabolism (1984; Mader and Heck, 1986; Mader, 2003) provides a time efficient estimate an athletes MLSS (Quittmann et al., 2020), figures 6 and 7 highlight the potential perils of deriving the vLamax from an inappropriate test duration. For example, determining the vLamax from a test duration which is too long will underestimate the maximal glycolytic rate leading to higher calculated MLSS (Mader & Heck, 1986; Hauser et al., 2014; Wackerhage et al., 2023). Subsequently, the calculated MLSS will be above the athlete's physiological maximal metabolic steady state, pushing the athlete into the unsustainable severe exercise domain (Burnley & Jones, 2018). Additionally, underestimating the vLamax may lead to inappropriate training recommendations to enhance the athletes MLSS based on metabolic profiles (Quittmann et al., 2020; Wackerhage et al., 2023).

### 4.4 The Influence of Modality, Whole, Upper, and Lower Body Exercise on $\nu La_{max}$

Distinct differences in the amplitude of vLamax are observed dependent upon the exercise modality irrespective of test duration (Figure 5). Key factors influencing vLamax include; total work completed (Quittmann et al., 2020), quantity of active muscle mass (Medbø & Toska, 2001; Quittmann et al., 2020; Quittmann et al., 2021b), intra-individual fibre type distribution between muscles (Johnson et al., 1973; Tesch et al., 1978), lactate distribution volume (Medbø & Toska, 2001), sport specific motor recruitment patterns (Quittmann et al., 2020; Quittmann et al., 2021b), and the participants modality specific training background (Held et al., 2024).

amplitude of vLa<sub>max</sub> has demonstrated to be higher, with more work achieved for a given duration, and associated with the amount of active muscle mass (Quittmann et al., 2020). The greater the active muscle mass results in an increased vLamax, when comparing exercise modalities utilising upper or lower body extremities in isolation (Figure 5; Quittmann et al., 2020; Quittmann et al., 2021b). Whilst our results illustrate the highest vLamax amplitude following cycling (Figure 5), Quittmann et al. (2021b) observed running elicits a higher vLamax compared with cycling in the same cohort of participants. Differences may be observed due to elite sprint populations who possess quantities of muscle mass around the quadriceps and hamstrings along with greater muscle pennation angles (Kordi et al., 2019) in comparison to endurance trained triathletes utilised by Quittmann et al. (2021b). Running may elicit higher vLamax in participants of the same cohort due to greater activation of knee flexors and hip extensors, compared with cycling, due to an increase in active muscle mass (Jönhagen et al., 1996; Dorel et al. 2012). Additionally, whilst Quittmann et al. (2020) controlled the cycling frequency at 130 rpm, during sprint running stride frequency can exceed 250 steps per minute (Morin et al., 2011) with a higher movement velocity associated with higher vLamax values (Haase et al., 2024).

Over a 15s test duration vLamax values for upper or lower body modalities correspond with the amount of active muscle mass; cycling ~ 25% body mass (Medbø & Toska, 2001), Kayak estimated higher than hand cycling (Forbes & Chilibeck, 2007), hand cycling ~ 6.5% body mass (estimated from Quittmann et al., 2021a), Knee extensor isokinetic force ~ 4% body mass (Bangsbo et al., 1990). Whilst the upper extremities have a higher percentage of fast-twitch muscle fibres compared to the lower (Johnson et al., 1973), higher muscle

cross sectional area and lean segmental mass of the lower body (Quittmann et al., 2021b) appear more influential in vLamax amplitude.

Intuitively whole-body exercise engaging larger volumes of muscle mass might be expected to elicit the highest amplitudes. However, during whole-body exercise the influence of the active muscle mass appears to be masked (Figure 5; Table 1). Low vLamax values observed in rowing maybe due to lower movement frequencies (Held et al., 2024). In rowing stroke rate peaks at ~50 strokes per minute (Held et al., 2024), in comparison to running, cycling, and handcycling vLamax tests are conducted at considerably higher movement frequencies >100 steps or rotations per minute. Haase et al. (2024) recently reported higher movement frequencies within cycling elicit higher vLamax amplitudes. This point has been supported by the recommendation Henneman's hierarchical size principle should incorporate movement velocity, with faster muscle fibre types being recruited with rising intensity and velocity (Dunst et al., 2024). Whilst the amplitude of vLamax was higher for swimming comparable to cycling and running for the same test duration, a higher vLamax was not attained despite the utilisation of larger quantities of muscle mass (Figure 5, Table 1). These values may also be observed due to slower movement frequencies, with the mean stroke rate within the 50m freestyle for males (59.58  $\pm$  5.40 cycles·min-1) and females (56.92 ± 3.24 cycles·min<sup>-1</sup>) reported by Pelayo et al. (1996). The findings from this systematic review suggest an augmented demand on anaerobic glycolysis and/or reduced lactate clearance with faster movement frequencies.

Lactate uptake of the upper extremities has been shown to be negligible during cycling (Medbø & Toska, 2001). During whole body exercise, despite high rates of lactate appearance, arterial blood lactate concentrations can be maintained, due to the large capacity of active skeletal muscle to consume lactate (van Hall et al., 2003). Furthermore, higher lactate uptake compared to release from the legs was shown to be, the primary factor for maintaining low arterial BLC (van Hall et al., 2003). Van Hall et al. (2003) demonstrated that only active skeletal muscle consumes large quantities of lactate during exercise, due to enhanced energy expenditure, and correlated with enhanced muscle blood flow. The exercise modality may influence the lactate volume distribution during and post exercise dependent upon changes in blood flow redistribution (Medbø & Toska, 2001). For example, Quittmann et al. (2021b) observed a higher lactate removal velocity following handcycling compared with cycling. This is potentially due to venous-arterial lactate differences being greater following upper vs lower body exercise (Klausen et al., 1974). Higher lactate uptake of the legs (van Hall et al., 2003), coupled with changes in lactate distribution volumes (Medbø & Toska, 2001), may explain why smaller amplitudes in vLamax are attained following whole-body vs lower body exercise.

In summary, distinct differences observed in the peak amplitude of vLamax between exercise modalities. These findings coupled with Quittmann et al. (2020; 2021a) comparisons highlight between modalities should not be drawn. Large interindividual differences, and low correlations between modalities (Quittmann et al. 2020; 2021a) demonstrate vLamax is modality specific, sensitive to training history, movement velocity, and extremity specific. Therefore, testing the vLamax should be specific to the sport modality (Held et al., 2024; Quittmann et al. 2020; 2021a).

### 4.5 vLa<sub>max</sub> Reliability

Reliability studies focusing on test duration currently make it difficult to assess the stability of the vLamax determination in the test-retest design. The available studies show moderate to excellent reliabilities based on correlation coefficients using a selected test duration. The test durations ranging from 8 to  $23.1 \pm 0.31$  s (Table 2). Only Wawer et al. (2020) compared the reliability of the vLamax across different test durations during running (non-motorized treadmill, running track), and found good to excellent reliabilities (ICC: 0.82 to 0.92). Lower correlation coefficients for test-retest trials were found by Nitzsche et al. (2018b) with isokinetic force loads. The coefficients were 0.72 at approximately 12 s and 0.48 at approximately 23 s, suggesting a lower reliability of the vLamax with increasing test duration. The BLC prior to the test (BLC<sub>pre</sub>) and the peak BLC (BLCpeak) are important in the calculation of vLamax and thus, their reliability should be considered. In the studies included, coefficients of correlation the demonstrated a large range of 0.3 to 0.8 (Quittmann et al. 2021b; Adam et al. 2015) and coefficient of variation up to 45.6 % (Harnish et al. 2023; Adam et al. 2015), which indicates high variability or low stability.

A recent study by Pohl et al. (2024) indicates a reduced  $\nu L_{amax}$  with higher BLC<sub>pre</sub>. The authors therefore suggest controlling BLC<sub>pre</sub> more carefully with a BLC<sub>pre</sub>  $\leq 1.5$  mmol  $\cdot$  L<sup>-1</sup> following the warm-up. Research by Wittekind and Beneke (2011) also found reduced glycolytic rate with increased BLC<sub>pre</sub> induced by a heavy warm-up. In this study a BLC<sub>pre</sub> of 2.0  $\pm$  0.3 mmol  $\cdot$  L<sup>-1</sup> did not significantly influence  $\Delta$ BLC. Since in both studies the high-BLC<sub>pre</sub>-condition were well above 1.5 and 2.0 mmol L<sup>-1</sup> (3.37  $\pm$  0.54 and 4.2  $\pm$  0.9 mmol  $\cdot$  L<sup>-1</sup>), it is still unknown at which concentration  $\Delta$ BLC is impaired. Irrespective of this, a light

warm-up with a control of BLC<sub>pre</sub> should be applied. A BLC<sub>pre</sub> of  $\leq 2.0$  or  $\leq 1.5$  mmol  $\cdot$  L<sup>-1</sup> can be assumed to not influence the  $\nu$ La<sub>max</sub>. This could also help to reduce the variability and reliability of BLC<sub>pre</sub>.

Furthermore, the correlation coefficients of the BLC<sub>peak</sub> range from 0.69 to 0.91 (Nitzsche et al. 2018b; Adam et al. 2015; Quittmann et al. 2021b; Held et al. 2024; Wawer et al. 2020) with coefficient of variation from 5.2 to 23.3% (Adam et al. 2015; Harnish et al. 2023; Wawer et al. 2020) and indicate moderate to good reliability. Studies that reported correlation coefficients for  $\Delta$ BLC reported values of 0.78 to 0.91 (Adam et al. 2015; Quittmann et al. 2021b; Meixner et al. 2024), which indicates good to excellent reliability. Although the reliability BLC<sub>pre</sub> was lower than BLC<sub>peak</sub>, this did not appear to affect the high stability of the  $\Delta$ BLC.

The largest degree of error that most significantly influences the reliability of the vLa<sub>max</sub> determination, is the alactic time span. Meixner et al. (2024), found that vLamax calculated using different methods of talac determination, showed considerable differences in correlation coefficients. As shown in Table 2, the highest coefficients were calculated for fixed talac (3.5 s). performance-dependent talac (TTP, TTP-3.5%) increase the standard error of measurement and coefficient of variance. Furthermore, Nitzsche et al. (2018b) showed decreasing correlation coefficients (<0.5) and increasing measurement errors for talac with increasing exercise time. The correlation coefficients of Wawer et al. (2020) appear to fluctuate unsystematically depending on the test duration (0.11-0.76). Only Adam et al. (2015) reports coefficients of performance-dependent talac (TTP-3.5%) greater than 0.8.

Lower reliability observed by Harnish et al. (2023) could stem from the use of a handheld blood lactate analyser (Lactate Plus) and an

unreported standardization of pedaling frequencies and load factor. The Lactate Plus samples a small quantity of blood (3-µl) and has been reported to produce a mean relative difference of -7% (2.5 to 97.5% percentile ranges -23 to 10%) when compared with the Biosen (Mentzoni et al., 2024). To standardise movement velocity vLamax tests are typically conducted in isokinetic mode (120, 130 rpm) on a cycle ergometer (Hauser et al., 2014, Adam et al., 2015, Quittmann et al., 2018, Quittmann et al., 2021a, Yang et al., 2023, Dunst et al., 2023a, b, Meixner et al., 2024, Poffé et al., 2024). Studies investigating the reliability of the vLamax as a function of movement frequency (cadence, angular velocity) are still pending, but it can be assumed that the vLamax is influenced by the movement velocity (Haase et al., 2024). The present reliability studies used only two repeated measurements (Harnish et al., 2023; Held et al. 2024; Meixner et al., 2024; Nitzsche et al., 2018b; Quittmann et al., 2021a; Quittmann et al., 2021b; Wawer et al., 2020) except Adam et al. (2015) who used three repeated trials.

Accurate comparisons in the reliability of between different populations including sex and performance level cannot be drawn. Seven of the eight studies examining reliability included both male (n = 196) and female (n = 65) participants (n = 261) (Table 2). However, no study directly compared the reliability of the vLamax or lactate kinetics between sexes. Whilst Harnish et al. (2023) observed no statistical differences in blood lactate parameters, alactic time, or vLamax between sexes, several studies report sex differences between both lactate parameters and vLamax (Adam et al., 2015; Held et al., 2024; Meixner et al., 2024b). Sex differences in the vLamax are likely to occur, with females likely to express lower maximal glycolytic rates due to differences in lean body mass (Miller et al., 2024), lower activities of PFK, pyruvate kinase,

lactate dehydrogenase, hexokinase, glycogenolysis phosphorylase, and succinic dehydrogenase (Green et al., 1984; Jaworowski et al., 2002). These findings suggest further exploration of sex differences are required. Due to the homogeneity of the participants performance level (Amateur/ University Students: n = 7; National Level: n = 1) comparisons in the test reliability cannot be drawn.

In summary, the current reliability of vLa<sub>max</sub> ranges from moderate to excellent (Table 2). The reliability of the vLa<sub>max</sub> can be enhanced by applying a fixed alactic timespan (Meixner et al. 2024), employing shorter test durations (Nitzsche et al., 2018b), standardising movement velocity where possible (Haase et al., 2024), and by controlling the BLC prior to the test within a narrow margin (Adam et al. 2015; Harnish et al. 2023; Pohl et al., 2024).

### 4.6 Training Interventions Influence on vLamax

It is assumed that a high vLamax is unfavorable in endurance disciplines. A higher νLamax with a constant VO2max leads to a reduction in performance at the MLSS (Wackerhage et al., 2022). This is evident, in lower  $\nu La_{max}$  values of 800 m runners compared to 100 m sprinters (Thron et al., 2024). In contrast, a high vLamax is necessary for sprint disciplines (Wackerhage et al., 2022). It seems apparent that highly endurance trained athletes have a lower vLamax compared to anaerobically trained athletes such as track cyclists (e.g. results from Yang et al. (2023) vs. Quittmann et al. (2021a)). At what point the vLamax is low or high is difficult to assess due to methodological differences determination. In addition, the vLamax and its adaptation should be considered combination with changes in VO2max and always with a view to the desired performance. Olbrecht and Mader (2005)describe improvements of the performance (e.g., shift of a lactate curve) can be the results of different combinations and adaptations of  $vLa_{max}$  ( $\uparrow \rightarrow \downarrow$ ) and  $\dot{V}O_{2max}$  ( $\uparrow \rightarrow \downarrow$ ). In line with this, Hauser et al. (2014) could show that the achieved power at the MLSS can almost be identical, but resultant from different combinations of  $vLa_{max}$  and  $\dot{V}O_{2max}$ . It is hypothesised that endurance training leads to a reduction in  $vLa_{max}$ , however, training intervention studies on the  $vLa_{max}$  are scarce. Where, only two studies met our inclusion criteria (Hommel et al., 2019; Nitzsche et al., 2020).

Endurance training methods, such moderate intensity continuous training or high intensity interval training typically result in an increase of VO<sub>2max</sub> (Poon et al., 2021), probably accompanied by a reduction of vLamax or no change of vLamax (Mader & Heck, 1986; Wackerhage et al., 2022). This hypothesis is supported by Hommel et al. (2019) who observed a significant reduction in vLamax and an increase in VO<sub>2max</sub> following 6-weeks of sprint interval training (30-40 mins with 4-6 Wingate)tests lasting 30 s each) in physical education students, who were not specifically trained in endurance or sprinting. High volume training (60 min at 1.5 to 2.5 mmol · L-1 BLC), on the other hand, increased VO<sub>2max</sub> but did not significantly alter vLamax (Hommel et al., 2019). However, the training interventions lacked standardisation of the intensity of both the sprint interval and highvolume training. Contrary, resistance training over 6-weeks resulted in an increase of vLamax regardless of intensity or training volume (Nitzsche et al., 2020). Furthermore, Sperlich et al. (2010) observed a significant increase of vLamax following 5-weeks of HIIT and a decrease following 5-weeks of high-volume training in swimming, contrary to Hommel et al. (2019). However, the vLamax test duration was ~85 s, well above the recommended test duration, and it is questionable whether the adaptation of the vLa<sub>max</sub> will be similar with a shorter test duration. An observational study by Manunzio et al. (2016) was conducted over six months prior to an ultraendurance race.  $\dot{V}O_{2max}$  increased moderately and  $vLa_{max}$  decreased significantly associated with an increased training volume around the MLSS and above, while the volume of low intensity training was reduced. The studies by Sperlich et al. (2010) and Manunzio et al. (2016) are both worth mentioning. However, both studies were excluded from the systematic review due to methodological limitations (test duration ~85 s) or due to the observational character of the study.

Data available does not allow any clear conclusions to be drawn. It is also unclear how the training status affects the adaptation of the vLamax. Further training studies are required in which intensity and volume are varied in small steps and the adaptations of vLamax and VO2max should be monitored carefully. Standardised methodologies for testing the vLamax needs to be agreed and applied to allow accurate comparisons to be drawn (See 4.8. for more details).

### 4.7 Blood Lactate Sampling

The most common blood lactate sampling method was sampling every minute for 10minutes post exercise for a total of 11 post exercise samples (Table 1). This was used in 17 of the 27 articles, with three other articles using similar methodology with additional samples. Exercise duration has a significant role in tBLC<sub>max</sub> whereby shorter tests durations lend themselves to a reduced tBLCmax. Langley et al. (2024) demonstrated tBLCmax peaked between  $5 \pm 2$  min,  $6 \pm 2$  mins, and  $7 \pm 2$  mins following 10, 15, and 30 s cycling durations. Mavroudi et al. (2023) also reported that the shorter the swimming distance the quicker the tBLC<sub>max</sub> in 25, 35, and 50 m swimming sprints, respectively. Therefore, the consensus should not be limited to a specific sampling duration post exercise, but be reflective of the environment the test is conducted in. If a vLamax is conducted in the field with applied practitioners, post exercise samples should be taken every minute (including immediately post exercise) until BLC $_{max}$  values are identified minus 1 mmol  $\cdot$  L $^{-1}$ . This will certify a true BLC $_{max}$  is detected whilst minimising the amount of post exercise samples. This maintains the integrity of the test whilst reducing the burden on the practitioner and the athlete.

In order to increase the accuracy of the BLC<sub>max</sub> determination and enforce strict scientific rigor, post exercise samples could be taken every 30 s (including immediately post exercise) until peak blood lactate values are identified minus 1 mmol·L-1. It could be argued this frequency of sampling would further improve accuracy. However, a study by Nitzsche & Haase (2023) showed no significant differences in the determination of BLCmax using a bi-exponential function, with a blood sampling rate every 30 s compared to every 60 s up to the 9th minute following a maximal sprint. The use of a bi-exponential function also offers the advantage of determining lactate kinetics (lactate invasion and elimination) more precisely. This is a growing interest in vLamax research but requires a longer post exercise measurement period (up to 30 minutes), thus more blood samples, whereby the frequency of the blood samples is reduced after the BLC<sub>max</sub> occurs (Beneke et al., 2010). Further, reporting tBLC<sub>max</sub> in future vLa<sub>max</sub> research will better inform the readers of the post exercise lactate kinetics across different modalities and populations.

Following the sprint, the athlete should remain inactive to not influence the BLC, as active recovery is known to increase the lactate elimination out of the blood compartment for example following multiple sprints (Kappenstein et al., 2015) and following a maximal incremental test (Taoutaou et al.,

1996). BLC<sub>peak</sub> was not influenced in these studies. However, it is currently unknown how active recovery affects the BLC after a single sprint, and a reduced BLC<sub>peak</sub> would result in reduced  $\Delta$ BLC and  $\nu$ La<sub>max</sub>. Researchers should clearly indicate whether their athletes/participants are active or passive after the sprint test.

### 4.8 Test Recommendations

### 4.8.1 General Test Recommendations

To allow accurate, reliable, and valid vLa<sub>max</sub> test results the protocol needs to be standardised specific to the test scenario and modality. To gain an accurate assessment during maximal performance tests the athlete should be familiarised with the test protocol (Ozkaya, 2013) and highly motivated to attain maximal effort (Rendos et al., 2019). Additionally, factors including baseline blood lactate concentration, and prior carbohydrate ingestion (Pohl et al., 2024) need to be considered to allow accurate and reliable results. Therefore, several general test recommendations should be implemented:

- 1) Baseline blood lactate should be < 1.5 mmol · L-¹ (Pohl et al., 2024).
- 2) Participants should consume a carbohydrate rich diet of >6g·kg body mass per day for 48-72 hours prior and avoid long and vigorous exercise 48-hours prior to ensure the participants attend in a glycogen rich state (Hawley et al., 1997).
- 3) Avoid consumption of any glucose containing beverages (e.g., juice, soft drinks) or energy gels in the hour prior to vLa<sub>max</sub> testing (Pohl et al., 2024).
- 4) Abstinence of caffeine (in any form) 6 12 hours prior. This recommendation stems from the potential influence of caffeine on the test results. Caffeine has been shown to increase the mean and peak power in a Wingate test (Grgic, 2018) and to increase the

BLC in submaximal exercise (Glaister & Gissane, 2018). Therefore, Glaister and Gissane (2018) emphasise the importance of caffeine withdrawal before any experimental intervention. However, withdrawal from caffeine can result in symptoms like headaches, tiredness/fatigue or decreased energy/activeness (Juliano & Griffiths, 2004), which may add a confounding variable (Merrell et al., 2024). The half-life of caffeine is generally 4 – 6 h but varies between 2 h to 10 h (Blanchard & Sawers, 1983; Guest et al., 2021) and caffeine withdrawal symptoms emerge after 12-24 h (Juliano & Griffiths, 2004). If caffeine abstinence is not possible, the participants are encouraged to document caffeine consumption and replicate consumption prior to all trials. This is particularly important repeated for measurements (e.g., in reliability or training studies).

- 5) Arrive in euhydrated state due to dehydration causing higher BLC during exercise (Green et al., 2018; Logan-Sprenger et al., 2013). To ensure adequate hydration it is recommended to consume a minimum of 40 mL·kg body mass of water the day prior (Vivanti, 2020).
- Participants should complete at least one allout familiarisation trial prior (Ozkaya, 2013).

- 7) Loud verbal encouragement should be provided throughout test procedures (Rendos et al., 2019).
- 8) vLa<sub>max</sub> test duration should be between 8 12 s irrespective of modality (Figure 5). Activities which are more conducive to assessment over a given distance e.g., running and swimming would benefit from aiming for a task completion time within this range.
- A passive seated rest period following the test during blood lactate sampling to minimise removal of lactate from the blood for metabolism.

### 4.8.2 vLa<sub>max</sub> Test Recommendations & Considerations

Currently, body the of research demonstrates an overwhelming lack consensus to testing the vLamax. A standardised test protocol must be implemented between scientists and practitioners to enhance the reliability of the results and allow for meaningful comparisons between data to be drawn. Table 4 outlines a standardised vLamax test protocol we advocate to be implemented between researchers and practitioners to provide continuity in measures derived from the results of this review.

**Table 4.** Recommendations for a standardised vLamax Test Protocol.

Time	Component	Recommended procedure				
Pre-Test	Nutritional Status	Glycogen rich state and Euhydrated: > 6g · kg body mass of CHO and 40 mL·kg body mass of water the day prior.				
	Baseline blood lactate	< 1.5 mmol · L <sup>-1</sup>				
	Test Duration	10 s				
Test	Alactic Time	3 s				
rest	Movement Velocity	Isokinetic mode (where applicable): Cycling cadence 130 rpm.				
	Encouragement	Loud vocal encouragement throughout.				
	Recovery	Seated passive recovery				
	Blood lactate sampling	BLC sampled from the earlobe every 60 s until BLC peak minus 1 mmol $\cdot$ L-1.				
Post-Test	Identification of BLC <sub>peak</sub>	Highest measured BLC.				
	vLa <sub>max</sub> formula	$vLa_{max}rac{BLa_{maxpost}-BLa_{pre}}{t_{test}-t_{alac}}$				

Scientific research aiming derive understanding mechanistic of energy metabolism are encouraged to adopt Yang et al. (2023) 'pure vLamax' formula to deduce the oxidative contribution from the test time. This method requires a metabolic cart to measure VO<sub>2</sub> before, during, and after, the νLamax test protocol to allow energy contributions to be derived from PCr-La-O2 energy equivalence (Yang et al., 2023). Whilst this approach provides valuable insight into the athletes underpinning metabolism, adoption of this procedure has been omitted from our recommendations due to the requirements of an expensive metabolic cart and additional mathematical procedures which complicated test procedures. Furthermore, the influence of the oxidative metabolism on the vLamax appears small when utilising a 10 s sprint (Langley et al., 2024; Langley & Porter, 2024).

If the researcher wishes to explore the mechanistic underpinnings of energy metabolism we advocate estimating the participants fat-free mass in accordance with Meixner et al. (2024). Estimation of the participants fat-free mass enhances the accuracy of determining the glycolytic energy contribution (Meixner et al., 2024b). The mechanical energy equivalent of 1 mmol · L<sup>-1</sup> of blood lactate accumulation can be calculated as 12 J/kg of fat-free mass (Meixner et al., 2024b).

To derive the most accurate determination of BLC<sub>max</sub> and subsequent vLa<sub>max</sub> researchers should employ a bi-exponential function as detailed by Beneke et al. (2005) and Beneke et al. (2007). As described in section 4.7. biexponential modelling provides further insight into blood lactate kinetics including lactate invasion and elimination time parameters, this approach has been readily employed by several researchers to gain further understanding of the athlete physiological profile (Dunst et al., 2023a, 2023b; Haase et al.,

2024; Quittmann et al., 2018, 2020, 2021a, 2021b). Whilst this approach offers the most determination of the accurate  $\nu La_{max}$ differences between peak concentrations measures and modelled values are likely to be small and place additional time requirements for the participant. To employ ease of testing for both scientists and practitioners the use of bi-exponential modelling has not been suggested within the standardised recommendations.

### 4.8.3 Modality specific recommendations

During cycle ergometry the movement velocity has been demonstrated to significantly influence the  $\nu La_{max}$  amplitude with higher pedalling frequencies stimulating higher  $\nu La_{max}$  amplitudes (Haase et al., 2024). Subsequently, pedaling frequency needs to be controlled. We recommend the  $\nu La_{max}$  test should be conducted in isokinetic mode at a pedalling cadence of  $\geq 130$  rpm in accordance with Haase et al. (2024). To reduce the loss of force transmission and engage the hamstring muscles in the upcycle of the pedal stroke participants should utilise clipless pedals where possible (Hintzy et al., 1999).

Currently, there is limited data to draw distinct sports specific  $\nu La_{max}$ recommendations for other sporting modalities. Only one study in running has been conducted to test the reliability between nonmotorised treadmill and track running (Wawer et al., 2020). Furthermore, within running no studies have evaluated the differences in vLamax tested over a set duration and set distance. Set running distances comparisons between individuals and studies challenging due to the time dependence of the vLa<sub>max</sub> formula. Swimming stroke frequency is likely to influence vLamax and therefore the vLamax is likely to be stroke specific due to differences in movement frequencies between strokes (Hellard et al., 2008). During isokinetic force testing the angular velocity influences the  $\nu La_{max}$  with  $210^{\circ} \cdot s^{-1}$  previously utilised as demonstrated to elicit the highest  $\nu La_{max}$  (Nitzsche et al., 2018b).

#### 4.9 Limitations

### 4.9.1 Limitations of Bi-exponential modelling to identify optimal test duration

Bi-exponential modelling time to peak vLamax from pooled data is not without limitations. Firstly, this data combines data from both sexes, varying training status (recreational – elite), and athletes with different specialisms (sprint or endurance). vLamax has been reported lower in females than their male counterparts (Harnish et al., 2023; Meixner et al., 2024b; Poffé et al., 2024, Thron et al., 2024) and inclusion of both sexes may lower the average peak vLamax, skewing results when compared to all male counterparts. Studies investigating the vLamax in elite level sprint cyclists (Dunst et al., 2023a, 2003b; Yang et al., 2023) report very high values, compared with elite endurance cyclists (Manunzio et al., 2016), and amateur cyclists (Harnish et al., 2023; Meixner et al., 2024b). Furthermore, two studies (Haase et al., 2024; Langley et al., 2024) investigating a 10 s test duration both utilised high pedaling cadences (90 – 170,  $145 \pm 7$  rpm) which promote higher vLamax results (Haase et al., 2024).

### 4.9.2 Standardisation of analysis of blood lactate accumulation kinetics

Limited studies (5) report the post exercise blood lactate kinetics including the time to peak or the rate of accumulation and rate of disappearance. Including post exercise blood lactate kinetics may allow for more accurate determination of when peak blood lactate occurs in varying populations and allows inferences of training status regarding lactate disappearance (Stallknecht et al., 1998). Furthermore, varying sampling times,

frequencies, and duration of post exercise blood lactate kinetics make comparisons of  $vLa_{max}$  values challenging. Whilst some studies calculate  $vLa_{max}$  from measured BLC others utilise bi-exponential modelling of blood lactate to identify peak concentrations. These differences in methods may influence the derived  $vLa_{max}$  making comparisons between studies challenging.

### 4.9.3 Standardisation of analysis of blood lactate accumulation kinetics

Inferences have been made between and within modalities in this review, although it should be outlined that movement velocities within modality can lead to distinct changes in vLamax. Movement velocities have been shown to be influential on lactate kinetics and energy availability (Gottshall et al., 1996, Gentil et al., 2006). Therefore, future research needs to consider how combining modalities via time under tension or speed of cyclical movement (rpm or cadence) may affect lactate kinetics thus vLamax.

### 4.9.4 Demographic of participants

Due to the moderate number of cycling studies, inferences from both elite and amateur populations were amalgamated. Cycling specific considerations in this review has consciously taken ability level into account. Future research is needed in a variety of populations (amateur, well-trained, & elite) in each modality (cycling & running) to ensure population specific inferences can be made. Additionally, inferences between endurance and sprint trained individuals were also combined due to the limited number of studies. Both limitations (participation level and training characteristic) are minor but require future research before direct comparisons can be made.

### 4.9.5 Limitations of the review process

The moderate number of articles included into some categories of this systematic review limits our ability to draw firm conclusions about the modality (swimming, rowing, and kayak) and duration specific inferences during a vLamax test. This further highlights the need for more studies of varying modalities and test durations in the vLamax literature. The limited number of training intervention studies leads to reduced inferences regarding if the vLamax is trainable and if so, what training interventions offer the best avenue to enhance or reduce the vLamax. Additionally, it can be noted that even though this systematic review followed the PRISMA guidelines, it is conceivable that some articles could have been missed or incorrectly excluded during the screening process.

The maximum lactate accumulation rate could also have clinical relevance in the future. Various diseases that affect muscle metabolism and cause maladaptation's in energy supply show a muscle fiber shift because of these changes (COPD, heart failure, type II diabetes). The vLamax/vLapeak could also be a significant factor in muscle diseases with genetically triggered enzyme defects that lead to faster exhaustion during physical activity due to disruption of muscle energy metabolism. Furthermore, it's possible the use the vLamax can be used as a metabolic performance parameter to determine muscle adaptations following training interventions as a minimal invasive method, as shown in an initial study (Nitzsche et al., 2021).

### 4.10 Future Research Directions

Based on the current body of evidence the key priority is to standardise the vLa<sub>max</sub> testing procedures within the scientific community and between practitioners. Therefore, we advocate for scientists and practitioners to

embrace the recommended procedures including:

- 1) A standardised test duration of 10s.
- 2) A standardised alactic time interval of 3 s.
- 3) Baseline blood lactate values below 1.5 mmol·L<sup>-1</sup>.
- 4) Post test blood lactate sampling every 60 s from the earlobe until a decline in 1 mmol · L<sup>-1</sup> from peak blood lactate is observed.
- 5) Standard passive seated recovery post sprint.

This systematic review has highlighted numerous gaps in the  $\nu La_{max}$  literature that need addressing to improve the rigor of the  $\nu La_{max}$  principle. We recommend that future research focuses on the following areas:

- 1) Currently, there are no validation studies analysing the vLa<sub>max</sub> measured from blood lactate and the *V*max of glycolysis. Therefore, validation studies are required to analyse the relationship between the maximal rate of glycolysis within the muscle and the vLa<sub>max</sub> assessed at the whole-body level.
- 2) Identification of which method offers the highest validity and reliability to estimate the alactic time interval.
- 3) Establish normative data for the  $\nu$ Lamax for both male and female populations.
- 4) Further analysis of how different training interventions influences the interactions between  $\nu La_{max}$ ,  $\dot{V}O_{2max}$ , MLSS and subsequent influence on performance.
- 5) Analysis of what variables impact predictions of the maximal metabolic steady state when applying Mader's mathematical model of human energy metabolism (Mader, 1984) and what approaches identify the most accurate model to determine the MLSS.

### 5 Conclusions

This systematic review of twenty-seven experimental research articles, outlines the current methodological approaches, provides recommendations to optimise vLa<sub>max</sub> procedures and future research. We also provide clarity that the vLa<sub>max</sub> formula derived from blood lactate concentrations was initially proposed by Mader (1994), with earlier mathematical models of human energy metabolism (Mader, 1984; Mader and Heck, 1986) applying maximal lactate formation rate directly within the muscle cell.

Identification of the alactic time-period introduces the largest area of error within the reliability of the vLamax. To minimise such error a fixed alactic timespan should be employed. The vLamax is modality specific and movement velocity dependent, thus vLamax should be tested specific to the sporting modality. Based on the results from this review we propose a standardised test procedure to be adopted by researchers and practitioners. Key test recommendations include: Pre-test blood lactate values < 1.5 mmol·L<sup>-1</sup>, a test duration of 10 s, a fixed alactic timespan of 3 s, standardised movement velocity applicable), a passive recovery, and post exercise blood lactate sampling every minute until peak minus 1 mmol·L<sup>-1</sup>.

**Funding:** RH receives a scholarship from the Sächsische Aufbaubank (SAB) with funds from the European Social Fund Plus (ESF Plus) and the Free State of Saxony.

**Acknowledgments:** The authors would like to acknowledge the work of authors of all the manuscripts included into the systematic review.

**Conflicts of Interest:** No conflicts of interest, financial or otherwise, are declared by the authors.

**Author Contribution:** All authors (JL, RH, NN & MP) contributed to the conception and design of the review, screening of articles, analysis of data and interpretation of results, and the construction of the manuscript from draft to final approved manuscript.

### References

- Adam, J., Oehmichen, M., Oehmichen, E., Rother, J., Müller, U., Hauser, T., Schulz, H. (2015). Reliability of the calculated maximal lactate steady state in amateur cyclists. *Biology of sport*, 32(2), 97-102. doi: 10.5604/20831862.1134311
- Bangsbo, J., Gollnick, P., Graham, T., Juel, C., Kiens, B., Mizuno, M., Saltin, B. (1990). Anaerobic energy production and O2 deficit-debt relationship during exhaustive exercise in humans. *The Journal of physiology*, 422(1), 539-559. doi: 10.1113/jphysiol.1990.sp018000
- 3. Beneke, R., Pollmann, C., Bleif, I., Leithauser, R.M., and Hutler, M. (2002). How anaerobic is the Wingate anaerobic test for humans? *European Journal of Applied Physiology*. 87 (4-5), 388-392. doi: 10.1007/s00421-002-0622-4
- Beneke, R., Hutler, M., Jung, M., & Leithauser, R. M. (2005). Modeling the blood lactate kinetics at maximal short-term exercise conditions in children, adolescents, and adults. Journal of applied physiology, 99(2), 499-504. doi: 10.1152/japplphysiol.00062.2005
- Beneke, R., Jumah, M. D., & Leithäuser, R. M. (2007).
   Modelling the lactate response to short-term all out exercise. Dynamic medicine, 6, 1-7. doi: 10.1186/1476-5918-6-10
- Beneke, R., Wittekind, A., Mühling, M., Bleif, I., Leithäuser, R. (2010). Lactate response to Short term Exercise with elevated starting levels. *European Journal of Applied Physiology*. 110(1), 215-218. doi: 10.1007/s00421-010-1481-z
- 7. Blanchard, J., & Sawers, S. J. (1983). The absolute bioavailability of caffeine in man. European Journal of Clinical Pharmacology, 24(1), 93–98.
- 8. Burnley, M., & Jones, A. M. (2016). Power–duration relationship: Physiology, fatigue, and the limits of human performance. *European Journal of Sport Science*, *18*(1), 1–12. doi: 10.1080/17461391.2016.1249524
- Brooks, G., Dubouchaud, H., Brown, M., Sicurello, J., Butz, C. (1999). Role of mitochondrial lactate dehydrogenase and lactate oxidation in the intracellular lactate shuttle. *Proceedings of the National Academy of Sciences*, 96(3), 1129-1134. doi: 10.1073/pnas.96.3.1129
- Brooks, G. (2012). Bioenergetics of exercising humans. *Comprehensive Physiology*, 2(1), 537-562. doi: 10.1002/cphy.c110007
- 11. Brooks, G. (2018). The science and translation of lactate shuttle theory. *Cell metabolism*, 27(4), 757-785. doi: 10.1002/cphy.c110007

- Chung, Y., Sharman, R., Carlsen, R., Unger, S., Larson, D., & Jue, T. (1998). Metabolic fluctuation during a muscle contraction cycle. *American Journal* of *Physiology-Cell Physiology*, 274(3), C846-C852. doi: 10.1152/ajpcell.1998.274.3.C846
- Cornish-Bowden, A. (2015). One hundred years of Michaelis–Menten kinetics. *Perspectives in Science*, 4, 3-9. doi: 10.1016/j.pisc.2014.12.002
- Danforth, W. (1965). Activation of glycolytic pathway in muscle. In B. Chance, R. W. Estabrook, & J. R. Williamson (Eds.), Control of Energy Metabolism. Academic Press.
- 15. Dawson, M. (1983). "Phosphorus Metabolites and the Control of Glycolysis Studied by Nuclear Magnetic Resonance," in Biochemistry of Exercise. *International Series of Sport Sciences*. Editors J. Knuttgen, J. Vogel, and J. Poortmans (Champaign, Illinois: Human Kinetics Publishers, Inc.)
- Dobson, G., Yamamoto, E., & Hochachka, P. (1986).
   Phosphofructokinase control in muscle: nature and reversal of pH-dependent ATP inhibition. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 250(1), R71-R76. doi: 10.1152/ajpregu.1986.250.1.R71
- 17. Dorel, S., Guilhem, G., Couturier, A., Hug, F. (2012). Adjustment of muscle coordination during an allout sprint cycling task. *Medicine and Science in Sports and* Exercise (44) 2154–2164 doi: 10.1249/MSS.0b013e3182625423
- 18. Dunst, A., Hesse, C., Feldmann, A., Holmberg, H. (2023a). A novel approach to determining the Alactic time span in connection with assessment of the maximal rate of lactate accumulation in elite track cyclists. *International Journal of Sports Physiology and Performance*, 18(2), 157-163. doi: 10.1123/ijspp.2021-0464
- 19. Dunst, A., Manunzio, C., Feldmann, A., Hesse, C. (2023b). Applications of near-infrared spectroscopy in "anaerobic" diagnostics–SmO 2 kinetics reflect PCr dephosphorylation and correlate with maximal lactate accumulation and maximal pedalling rate. *Biology of Sport*, 40(4), 1019-1031. doi: 10.5114/biolsport.2023.122481
- Dunst, A., Hesse, C., Ueberschär, O. (2024). Understanding optimal cadence dynamics: a systematic analysis of the power-velocity relationship in track cyclists with increasing exercise intensity. Frontiers in Physiology, 15, 1343601. doi: 10.3389/fphys.2024.1343601
- Emhoff, C, Messonnier, L. (2023) Concepts of lactate metabolic clearance rate and lactate clamp for metabolic inquiry: a mini-review. *Nutrients*. 15(1) doi: 10.3390/nu15143213

- Esbjornsson-Liljedahl, M., Sundberg, C., Norman, B., Jansson, E. (1999). Metabolic response in type I and type II muscle fibers during a 30-s cycle sprint in men and women. *Journal of Applied Physiology*. 87(4), 1326-1332. doi: 10.1152/jappl.1999.87.4.1326
- Erecińska, M., Deas, J., & Silver, I. A. (1995). The effect of pH on glycolysis and phosphofructokinase activity in cultured cells and synaptosomes. *Journal of Neurochemistry*, 65(6), 2765-2772. doi: 10.1046/j.1471-4159.1995.65062765.x
- Ferguson, H. A., Zhou, T., Harnish, C., & Chase, J. G. (2021a). Model of 30-s sprint cycling performance: Don't forget the aerobic contribution!. IFAC-PapersOnLine, 54(15), 316-321. doi: 10.1016/j.ifacol.2021.10.275
- Ferguson, H. A., Harnish, C., & Chase, J. G. (2021b).
   Using field-based data to model sprint track cycling performance. Sports medicine-open, 7(1), 20. doi: 10.1186/s40798-021-00310-0
- Ferguson, H., Harnish, C., Klich, S., Michalik, K., Dunst, A. K., Zhou, T., & Chase, J. G. (2023). Powerduration relationship comparison in competition sprint cyclists from 1-s to 20-min. Sprint performance is more than just peak power. *Plos one*, 18(5), e0280658. doi: 10.1371/journal.pone.0280658
- 27. Ferretti, G. (2015). Energetics of muscular exercise. Springer International Publishing: Heidelberg. doi: 10.1007/978-3-319-05636-4
- 28. Forbes, S., Kennedy, M., Bell, G. (2013). Timemotion analysis, heart rate, and physiological characteristics of international canoe polo athletes. *The Journal of Strength & Conditioning Research*, 27(10), 2816-2822. doi: 10.1519/JSC.0b013e318280d2a2
- Freund, H., Zouloumian, P., Enguelle, S., Lampert,
   E. (1984). Lactate kinetics after maximal exercise in man. In Physiological chemistry of training and detraining (Vol. 17, pp. 9-24). Karger Publishers.
- Gentil, P., Oliveira, E., Bottaro, M. (2006) Time under Tension and Blood Lactate Response during Four Different Resistance Training Methods. *Journal of Physiological Anthropology*. 1(5) 339-344. doi: 10.2114/jpa2.25.339
- 31. Gottshall, R.W, Bauer, T.A., Fahrner S.L (1996) Cycling cadence alters exercise Hemodynamics. International Journal of Sports Medicine. 17(1): 17-21. doi: 10.1055/s-2007-972802
- 32. Glaister, M., & Gissane, C. (2018). Caffeine and Physiological Responses to Submaximal Exercise: A Meta-Analysis. International Journal of Sports Physiology and Performance, 13(4), 402–411. doi: 10.1123/ijspp.2017-0312

- Guest, N. S., VanDusseldorp, T. A., Nelson, M. T., Grgic, J., Schoenfeld, B. J., Jenkins, N. D. M., Arent, S. M., Antonio, J., Stout, J. R., Trexler, E. T., Smith-Ryan, A. E., Goldstein, E. R., Kalman, D. S., & Campbell, B. I. (2021). International society of sports nutrition position stand: Caffeine and exercise performance. Journal of the International Society of Sports Nutrition, 18(1), 1. doi: 10.1186/s12970-020-00383-4
- 34. Green, H. J., Fraser, I. G., & Ranney, D. A. (1984). Male and female differences in enzyme activities of energy metabolism in vastus lateralis muscle. Journal of the neurological sciences, 65(3), 323-331. doi: 10.1016/0022-510X(84)90095-9
- Green, J., Miller, B., Simpson, J., Dubroc, D., Keyes, A., Neal, K., Gann, J., Andre, T. (2018) Effects of 2% Dehydration on Lactate Concentration During Constant-Load Cycling. *Journal of Strength and Conditioning Research*. 32(7), 2066-2071. doi: 10.151910.1519/JSC.00000000000002293
- 36. Greenhaff, P., Nevill, M., Soderlund, K., Bodin, K., Boobis, L., Williams, C., Hultman, E. (1994). The metabolic responses of human type I and II muscle fibres during maximal treadmill sprinting. *The Journal of physiology*, 478(1), 149-155. doi: 10.1113/jphysiol.1994.sp020238
- Grgic, J. (2018). Caffeine ingestion enhances Wingate performance: A meta-analysis. European Journal of Sport Science, 18(2), 219–225. doi: 10.1080/17461391.2017.1394371
- Haase, R., Dunst, A., Nitzsche, N. (2024). The influence of pedaling frequency on blood lactate accumulation in cycling sprints. *International Journal* of Sports Medicine. 45(08): 608-615. doi: 10.1055/a-2255-5254
- 39. Hanon, C., Rabate, M., Thomas, C. (2011). Effect of Expertise on Postmaximal Long Sprint Blood Metabolite Responses. *Journal of Strength and Conditioning Research* 25(9): 2503-2509. | doi: 10.1519/JSC.0b013e3182001807
- 40. Harnish, C., Swensen, T., King, D. (2023) Reliability of the 15-s Maximal Lactate Accumulation Rate (VLamax) Test for Cycling. Physiologia 3(4): 542-551. doi: 10.3390/physiologia3040040
- Harris, R., Hultman, E., Nordesjö, L. (1974). Glycogen, Glycolytic Intermediates and High-Energy Phosphates Determined in Biopsy Samples of Musculus Quadriceps Femoris of Man at Rest. Methods and Variance of Values. Scandinavian journal of clinical and laboratory investigation, 33, 109– 120. doi: 10.3109/00365517409082477

- Hauser, T., Adam, J., Schulz, H. (2014). Comparison of calculated and experimental power in maximal lactate-steady state during cycling. *Theoretical* biology and medical modelling, 11, 1-12. doi: 10.1186/1742-4682-11-25
- 43. Hawley, J., Schabort, E., Noakes, T., Dennis, S. (1997). Carbohydrate-loading and exercise performance: an update. *Sports Medicine*, 24, 73-81. doi: 10.2165/00007256-199724020-00001
- 44. Heck, H., Schulz, H., Bartmus U. (2003). Diagnostics of anaerobic power and capacity. *European Journal of Sport Science*, 3: 1-23. doi: 10.1080/17461390300073302
- Held, S., Rappelt, L., Brockherde, J., Donath, L. (2024). Reliability of the Maximal Lactate Accumulation Rate in Rowers. *International Journal of Sports Medicine*. 45:238-244. doi: 10.1055/a-2206-4959
- Hellard, P., Dekerle, J., Avalos, M., Caudal, N., Knopp, M., & Hausswirth, C. (2008). Kinematic measures and stroke rate variability in elite female 200-m swimmers in the four swimming techniques: Athens 2004 Olympic semi-finalists and French National 2004 Championship semi-finalists. *Journal* of Sports Sciences, 26(1), 35-46. doi: 10.1080/02640410701332515
- Hintzy, F., Belli, A., Grappe, F., & Rouillon, J. D. (1999). Effet de l'utilisation de pédales automatiques sur les caractéristiques mécaniques mesurées lors de sprints sur cycloergomètre non isocinétique. *Science & sports*, 14(3), 137-144. doi: 10.1016/S0765-1597(99)80055-0
- Hirvonen, J., Rehunen, S., Rusko, H., & Härkönen, M. (1987). Breakdown of high-energy phosphate compounds and lactate accumulation during short supramaximal exercise. European Journal of Applied Physiology and Occupational Physiology, 56(3), 253– 259. doi: 10.1007/BF00690889
- Hirvonen, J., Nummela, A., Rusko, H., Rehunen, S., Härkönen, M. (1992). Fatigue and changes of ATP, creatine phosphate, and lactate during the 400-m sprint. The Canadian Journal of Sport Science, 17(2), 141-144. PMID: 1324108
- Hommel, J., Öhmichen, S., Rudolph, U., Hauser, T., Schulz, H. (2019). Effects of six-week sprint interval or endurance training on calculated power in maximal lactate steady state. Biology of Sport. 36: 47–54. doi: 10.5114/biolsport.2018.78906
- Jacobs, I., Tesch, P., Bar-Or, O., Karlsson, J., Dotan, R. (1983). Lactate in human skeletal muscle after 10 and 30 s of supramaximal exercise. *Journal of Applied Physiology*, 55(2), 365-367. doi: 10.1152/jappl.1983.55.2.365

- 52. Jaworowski, Å., Porter, M. M., Holmbäck, A. M., Downham, D., & Lexell, J. (2002). Enzyme activities in the tibialis anterior muscle of young moderately active men and women: relationship with body composition, muscle cross-sectional area and fibre type composition. Acta Physiologica Scandinavica, 176(3), 215-225. doi: 10.1046/j.1365-201X.2002.t01-2-01004.x
- 53. Jönhagen, S., Ericson, M., Nemeth, G., Eriksson, E. (1996). Amplitude and timing of electromyographic activity during sprinting. *Scandinavian Journal of Medicine and Science in Sports*, 6(1), 15-21. doi: 10.1111/j.1600-0838.1996.tb00064.x
- 54. Juliano, L. M., & Griffiths, R. R. (2004). A critical review of caffeine withdrawal: Empirical validation of symptoms and signs, incidence, severity, and associated features. Psychopharmacology, 176(1), 1–29. doi: 10.1007/s00213-004-2000-x
- 55. Kappenstein, J., Engel, F., Fernández-Fernández, J., & Ferrauti, A. (2015). Effects of active and passive recovery on blood lactate and blood pH after a repeated sprint protocol in children and adults. *Pediatric Exercise Science*, 27(1), 77–84. doi: 10.1123/pes.2013-0187
- Kavanagh, M., Jacobs, I. (1988). Breath-by-breath oxygen consumption during performance of the Wingate Test. Canadian journal of sport sciences 13(1), 91-93.
- 57. Klausen, K., Rasmussen, B., Clausen, J., Trap-Jensen, J. (1974). Blood lactate from exercising extremities before and after arm or leg training. *American Journal of Physiology-Legacy Content*, 227(1), 67-72. doi: 10.1152/ajplegacy.1974.227.1.67
- 58. Knuttgen, H. (1970). Oxygen debt after submaximal physical exercise. *Journal of Applied Physiology*. 29(5):651–657. doi: 10.1152/jappl.1970. 29.5.651
- Kordi, M., Folland, J., Goodall, S., Haralabidis, N., Maden-Wilkinson, T., Sarika Patel, T., ... & Howatson, G. (2020). Mechanical and morphological determinants of peak power output in elite cyclists. Scandinavian Journal of Medicine & Science in Sports, 30(2), 227-237. doi: 10.1111/sms.13570
- 60. Langley, J., Ng, S., Todd, E., Porter, M. (2024). VLamax: determining the optimal test duration for maximal lactate formation rate during all-out sprint cycle ergometry. *European Journal of Applied Physiology*, 1-12. doi: 10.1007/s00421-024-05456-9
- Langley, J.O., Porter, M.S. VLamax: determining the optimal test duration for maximal lactate formation rate during all-out sprint cycle ergometry. Response. Eur J Appl Physiol 124, 3149–3150 (2024). doi: 10.1007/s00421-024-05507-1

- 62. Lievens, E., Klass, M., Bex, T., & Derave, W. (2020). Muscle fiber typology substantially influences time to recover from high-intensity exercise. *Journal of applied physiology*, 128(3), 648-659. doi: 10.1152/japplphysiol.00636.2019
- Logan-Sprenger, H., Heigenhauser, G., Jones, G., Spriet, L. (2013) Increase in skeletal-muscle glycogenolysis and perceived exertion with progressive dehydration during cycling in hydrated men. *International journal of sport nutrition and exercise metabolism*. 23 (3), 220-229. doi: 10.1123/ijsnem.23.3.220
- 64. Mader, A. (1984). Eine Theorie zur Berechnung der Dynamik und des steady state von Phosphorylierungszustand und Stoffwechselaktivität der Muskelzelle als Folge des Energiebedarfs. na.
- 65. Mader, A., Heck, H. (1986). A theory of the metabolic origin of "anaerobic threshold". *International Journal of Sports Medicine*. 7: 45-65.
- 66. Mader, (1994).Aussagekraft der Laktatleistungskurve in Kombination mit Bestimmung anaeroben Tests zur der Stoffwechselkapazität. In Stellenwert der Laktatbestimmung in der Leistungsdiagnostik: 32 Tabellen; Clasing, D., Ed.; G. Fischer: Stuttgart, Germany pp. 133–152.
- 67. Mader A. (2003) Glycolysis and oxidative phosphorylation as a function of cytosolic phosphorylation state and power output of the muscle cell. European Journal of Applied Physiology. 88: 317-338, 2003. doi: 10.1007/s00421-002-0676-3
- 68. Mader, A. (2015). Die Chimäre des Dopings und die Irrealität der Trainingswissenschaft: Das deutsche Hochleistungssportsystem als Staatsreligion oder warum deutsche Sportler nicht mehr so erfolgreich sind, wie sie sein könnten (1. Aufl.). Buchwerkstatt.
- Margaria, R., Edwards, H., Dill, D. (1933). The possible mechanisms of contracting and paying the oxygen debt and the role of lactic acid in muscular contraction. *American Journal of Physiology-Legacy Content*, 106(3), 689-715. doi: 10.1152/ajplegacy.1933.106.3.689
- 70. Margaria, R., Aghemo, P., & Sassi, G. (1971). Lactic acid production in supramaximal exercise. *Pflügers Archiv: European Journal of Physiology*, 326, 152-161. doi: 10.1007/BF00586907
- 71. Mavroudi, M., Kabasakalis, A., Petridou, A., Mougios, V. (2023) Blood lactate and maximal lactate accumulation rate at three Sprint swimming distances in highly trained and elite swimmers. *Sports*, 11: 0-87. doi: 10.3390/sports11040087

- 72. Manunzio, C., Mester, J., Kaiser, W., Wahl, P. (2016). Training intensity distribution and changes in performance and physiology of a 2nd place finisher team of the race across America over a 6 month preparation period. *Frontiers in Physiology*, 7, 642. doi: 10.3389/fphys.2016.00642
- 73. Medbø, J., Toska, K. (2001). Lactate release, concentration in blood, and apparent distribution volume after intense bicycling. *The Japanese Journal of physiology*, 51(3), 303-312. doi: 10.2170/jiphysiol.51.303
- 74. Mentzoni, F., Skaugen, M., Eythorsdottir, I., Roterud, S., Johansen, E., Losnegard, T. (2024). Precision and accuracy of four handheld blood lactate analyzers across low to high exercise intensities. European Journal of Applied Physiology, 1-8. doi: 10.1007/s00421-024-05572-6
- 75. Merrell, L. H., Perkin, O. J., Bradshaw, L., Collier-Bain, H. D., Collins, A. J., Davies, S., Eddy, R., Hickman, J. A., Nicholas, A. P., Rees, D., Spellanzon, B., James, L. J., McKay, A. K. A., Smith, H. A., Turner, J. E., Koumanov, F., Maher, J., Thompson, D., Gonzalez, J. T., & Betts, J. A. (2024). Myths and Methodologies: Standardisation in Human Physiology Research-Should We Control the Controllables? International Journal of Sport Nutrition and Exercise Metabolism, 34(4), 242–250. doi: 10.1123/ijsnem.2024-0091
- 76. Michaelis, L., Menten, M. (1913). Die binetib der intertinwerbung. Biochem. Zeitung 49, 333-369.
- 77. Miller, R., Balshaw, T. G., Massey, G. J., Maeo, S., Lanza, M. B., Haug, B., ... & Folland, J. P. (2024). Sex differences in muscle morphology between male and female sprinters. Journal of Applied Physiology, 136(6), 1568-1579. doi: 10.1152/japplphysiol.00009.2023
- 78. Moher, D. (2009). Protocol: reporting guidelines systematic review. https://www.ohri.ca/protocols/RGSR\_Protocol.pdf
- 79. Morin, J. B., Bourdin, M., Edouard, P., Peyrot, N., Samozino, P., & Lacour, J. R. (2012). Mechanical determinants of 100-m sprint running performance. European journal of applied physiology, 112, 3921-3930. doi: 10.1007/s00421-012-2379-8
- McCully, K., Iotti, S., Kendrick, K., Wang, Z., Posner, J., Leigh Jr, J., Chance, B. (1994). Simultaneous in vivo measurements of HbO2 saturation and PCr kinetics after exercise in normal humans. *Journal of Applied Physiology*, 77(1), 5-10. doi: 10.1152/jappl.1994.77.1.5

- 81. Meixner, B., Nusser, V., Koehler, K., Sablain, M., Boone, J., Sperlich, B. (2024a) Reliability of power output, maximal rate of capillary blood lactate accumulation, and phosphagen contribution time following 15-s sprint cycling in amateur cyclists. *Physiological Reports*. 12(10): e16086. doi: 10.14814/phy2.16086
- Meixner, B. J., Nusser, V., Koehler, K., Sablain, M., Boone, J., & Sperlich, B. (2024b). Relationship of peak capillary blood lactate accumulation and body composition in determining the mechanical energy equivalent of lactate during sprint cycling. *European Journal of Applied Physiology*, 124(11), 3399-3407. doi: 10.1007/s00421-024-05529-9
- 83. Nitzsche, N., Zschäbitz, D., Baumgärtel, L., Schulz, H. (2017). Einfluss der Methode zur Bestimmung des alaktaziden Zeitintervalls bei isokinetischen Kraftbelastungen. In A. Schwirtz, F. Mess, Y. Demetriou, & V. Senner (Eds.), Schriften der Deutschen Vereinigung für Sportwissenschaft: Band 265. Innovation & Technologie im Sport: 23. dvs-Hochschultag, München, 13.-15. September 2017: Abstracts (p. 215). Feldhaus Edition Czwalina
- 84. Nitzsche, N., Baumgärtel, L., Schulz, H. (2018a). Comparison of maximum lactate formation rates in ergometer sprint and maximum strength loads. Deutsche Zeitschrift für Sportmedizin. 1: 13–18. doi: 10.5960/dzsm.2017.312
- Nitzsche, N., Baumgärtel, L., Maiwald, C., Schulz, H. (2018b) Reproducibility of Blood Lactate Concentration Rate under Isokinetic Force Loads. Sports (Basel). 20;6(4):150. doi: 10.3390/sports6040150
- Nitzsche, N., Lenz, J., Voronoi, P., Schulz, H. (2020) Adaption of Maximal Glycolysis Rate after Resistance Exercise with Different Volume Load. Sports Medicine Internation Open. 4: 39-44. doi: 10.1055/a-1146-4236
- 87. Nitzsche, N., Augustin, N., Klotz, M., & Schulz, H. (2021). Progressive exercise therapy in muscle dystrophy: two case studies in adult patients with DM2 and LGMD2D. *Internal Journal of Sports Medicine and Rehabilitation*, 4, 17. doi: 10.28933/ijsmr-2020-11-2605
- 88. Nitzsche, N., Haase, R. (2023). Einfluss des Abnahmeintervalls von Kapillarblutproben beim isokinetischen Radsprint auf die Laktatkinetik. In Association for the Promotion of Sports Medicine Hannover e.V. (Ed.), Sports Medicine and Health Summit 2023 - Scientific Abstracts. German Journal of Sports Medicine, 74(4), 103.

- 89. O'Brien, B., Payne, W., Gastin, P., Burge, C. (1997). A comparison of active and passive warm ups on energy system contribution and performance in moderate heat. *Australian Journal of Science and Medicine in Sport*, 29(4), 106-109.
- Olbrecht, J., & Mader, A. (2006). Individualization of training based on Metabolic Measures. In First International Symposium Sciences and practices in Swimming. Atlantica: Paris (pp. 109-115).
- 91. Ozkaya, O. (2013). Familiarization effects of an elliptical all-out test and the wingate test based on mechanical power indices. *Journal of Sports Science and Medicine*, 12(3), 521.
- 92. Park, S., Park, D., Kim, M., Lee, E., Lee, D., Jung, J., Yang, W. (2021). High-intensity warm-up increases anaerobic energy contribution during 100-m sprint. *Biology*, 10(3), 198. doi: 10.3390/biology10030198
- 93. Parolin, M., Chesley, A., Matsos, M., Spriet, L., Jones, N., Heigenhauser, G. (1999). Regulation of skeletal muscle glycogen phosphorylase and PDH during maximal intermittent exercise. *American Journal of Physiology-Endocrinology and Metabolism*. doi: 10.1152/ajpendo.1999.277.5.E890
- 94. Pelayo, P., Sidney, M., Kherif, T., Chollet, D., Tourny, C. (1996). Stroking characteristics in freestyle swimming and relationships with anthropometric characteristics. *Journal of applied biomechanics*, 12(2), 197-206. doi: 10.1123/jab.12.2.197
- 95. Pohl, A., Schunemann, F., Schaaf, K., Yang, W., Heck, H., Heine, O., Jacko, D., & Gehlert, S. (2024). Increased resting lactate levels and reduced carbohydrate intake cause vLamax underestimation by reducing the net lactate accumulation A pilot study in young adults. *Physiological Reports*, 12 (16), 1-16. doi: 10.14814/phy2.70020
- 96. Poffé, C., Van Dael, K., Van Schuylenbergh, R. (2024) INSCYD physiological performance software is valid to determine the maximal lactate steady state in male and female cyclists. Frontiers in Sports and Active Living. 6(1). doi: 10.3389/fspor.2024.1376876
- Poon, E. T.-C., Wongpipit, W., Ho, R. S.-T., & Wong, S. H.-S. (2021). Interval training versus moderateintensity continuous training for cardiorespiratory fitness improvements in middle-aged and older adults: A systematic review and meta-analysis. Journal of Sports Sciences, 39(17), 1996–2005. doi: 10.1080/02640414.2021.1912453
- Porter, M., & Langley, J. (2025). The relationship between muscle oxygen saturation kinetics and maximal blood lactate accumulation rate across varying sprint cycle durations. European Journal of Sport Science, 25(3), e12242. doi: 10.1002/ejsc.12242

- Quittmann, O., Abel, T., Zeller, S., Foitschik, T., Strüder, H. (2018). Lactate kinetics in handcycling under various exercise modalities and their relationship to performance measures in ablebodied participants. *European Journal of Applied Physiology*, 118: 1493-1505. doi: 10.1007/s00421-018-3879-y
- 100. Quittmann, O., Appelhans, D., Abel, T., Strüder, H. (2020) Evaluation of a sport-specific field test to determine maximal lactate accumulation rate and sprint performance parameters in running. *Journal of Science and Medicine in Sport*. 23: 27-34. doi: 10.1016/j.jsams.2019.08.013
- Quittmann, O., Abel, T., Vafa, R., Mester, J., Schwarz, Y., Strüder, H. (2021a) Maximal lactate accumulation rate and post-exercise lactate kinetics in handcycling and cycling. *European Journal of Sport Science*. 21: 539-551. doi: 10.1080/17461391.2020.1756420
- 102. Quittmann, O., Schwarz, Y., Mester, J., Foitschik, T., Abel, T., Strüder, H. (2021b) Maximal lactate accumulation rate in all-out exercise differs between cycling and running. *International Journal of Sports Medicine*. 42: 314-322. doi: 10.1055/a-1273-7589
- 103. Quittmann, O., Foitschik, T., Vafa, R., Freitag, F., Sparmann, N., Nolte, S., Abel, T. (2023) Is maximal lactate accumulation rate promising for improving 5000-m prediction in running?. *Internation Journal of Sports Medicine*, 44(04), 268-279. doi: 10.1055/a-1958-3876
- 104. Rendos, N., Harriell, K., Qazi, S., Regis, R., Alipio, T., Signorile, J. (2019). Variations in verbal encouragement modify isokinetic performance. *The Journal of Strength & Conditioning Research*, 33(3), 708-716. doi: 10.1519/JSC.00000000000002998
- 105. Robin, M., Nordez, A., Dorel, S. (2022). Analysis of elite road-cycling sprints in relation to maximal power-velocity-endurance profile: a longitudinal one-case study. *Scandinavian Journal of Medicine & Science in Sports*, 32(3), 598-611. doi: 10.1111/sms.14103
- 106. Sahlin, K., Harris, R., Nylind, B., Hultman, E. (1976) Lactate content and pH in muscle samples obtained after dynamic exercise. *Pflügers Archiv: European Journal of Physiology*. 367(1): 143-149, doi: 10.1007/BF00585150
- Rogatzki, M., Ferguson, B., Goodwin, M., Gladden,
   L. (2015). Lactate is always the end product of glycolysis. Frontiers in Neuroscience. 9: doi: 10.3389/fnins.2015.00022

- 108. Sašek, M., Mirkov, D. M., Hadžić, V., Šarabon, N. (2022). The validity of the 2-point method for assessing the force-velocity relationship of the knee flexors and knee extensors: the relevance of distant force-velocity testing. *Frontiers in Physiology*, 13, 849275. doi: 10.3389/fphys.2022.849275
- 109. Schünemann, F., Park, S., Wawer, C., Theis, C., Yang, W., Gehlert, S. (2023). Diagnostics of vLa.max and Glycolytic Energy Contribution Indicate Individual Characteristics of Anaerobic Glycolytic Energy Metabolism Contributing to Rowing Performance. *Metabolites*. 13(1). doi: 10.3390/metabol3030317
- 110. Smith, J., Hill, D. (1991). Contribution of energy systems during a Wingate power test. *British Journal of Sports Medicine*, 25(4), 196-199. doi: 10.1136/bjsm.25.4.196
- 111. Sperlich, B., Zinner, C., Heilemann, I., Kjendlie, P., Holmberg, H., Mester, J. (2010). High-intensity interval training improves VO2peak, maximal lactate accumulation, time trial and competition performance in 9-11-year-old swimmers. *European Journal of Applied Physiology*. 110: 1029-1036. doi: 10.1007/s00421-010-1586-4
- 112. Stallknecht, B., Vissing, J., & Galbo, H. (1998). Lactate production and clearance in exercise. Effects of training. A mini-review. *Scandinavian journal of medicine & science in sports*, 8(3), 127-131. doi: 10.1111/j.1600-0838.1998.tb00181.x
- 113. Taoutaou, Z., Granier, P., Mercier, B., Mercier, J., Ahmaidi, S., & Prefaut, C. (1996). Lactate kinetics during passive and partially active recovery in endurance and sprint athletes. European Journal of Applied Physiology and Occupational Physiology, 73(5), 465–470. doi: 10.1007/BF00334425
- 114. Teixeira, C., Mezzaroba, P., Peserico, C., Machado, F. (2022). Effect of photobiomodulation on maximal lactate production rate on swimmers: a randomized, crossover, double-blind and placebo-controlled study. *Motriz: Revista de Educação Fisica*. 28: 2022. doi: 10.1590/S1980-6574202200017121
- 115. Tesch, P., Sjodin, B., Karlsson, J. (1978). Relationship between lactate accumulation, LDH activity, LDH isozyme and fibre type distribution in human skeletal muscle. *Acta Physiologica Scandinavica*, 103(1), 40-46. doi: 10.1111/j.1748-1716.1978.tb06188.x
- 116. Thron, M., Woll, A., Doller, L., Quittmann, O., Härtel, S., Ruf, L., Altmann, S. (2024) Physiological and Locomotor Profiling Enables to Differentiate Between Sprinters, 400-m Runners, and Middle-Distance Runners. *Journal of Strength and Conditioning Research*. 1;38(8):1419-1427. doi: 10.1519/JSC.000000000000004801

- 117. Van Hall, G., Jensen-Urstad, M., Rosdahl, H., Holmberg, H., Saltin, B., Calbet, J. (2003). Leg and arm lactate and substrate kinetics during exercise. *American journal of physiology-endocrinology and metabolism*, 284(1), E193-E205. doi: 10.1152/ajpendo.00273.2002
- 118. Van Hooren, B., Souren, T., Bongers, B. (2024). Accuracy of respiratory gas variables, substrate, and energy use from 15 CPET systems during simulated and human exercise. Scandinavian Journal of Medicine & Science in Sports, 34(1), e14490. doi: 10.1111/sms.14490
- 119. Vivanti, A. P. (2012). Origins for the estimations of water requirements in adults. *European Journal of Clinical Nutrition*, 66(12), 1282-1289. doi: 10.1038/ejcn.2012.157
- 120. Wackerhage, H., Hoffmann, U., Essfeld, D., Leyk, D., Mueller, K., & Zange, J. (1998). Recovery of free ADP, Pi, and free energy of ATP hydrolysis in human skeletal muscle. Journal of applied physiology, 85(6), 2140-2145. doi: 10.1152/jappl.1998.85.62140
- 121. Wackerhage, H., Gehlert, S., Schulz, H., Weber, S., Ring-Dimitriou, S., & Heine, O. (2022). Lactate Thresholds and the Simulation of Human Energy Metabolism: Contributions by the Cologne Sports Medicine Group in the 1970s and 1980s. *Frontiers in Physiology*, 13, 899670. doi: 10.3389/fphys.2022.899670
- 122. Wackerhage, H., Kabasakalis, A., Seiler, S., & Heck, H. (2025). Is the v Lamax for Glycolysis What the VO2 max is for Oxidative Phosphorylation? Sports Medicine, 1-14. doi: 10.1007/s40279-025-02259-6
- 123. Wawer, C., Heine, O., Predel, H., Park, D., Yang, W. (2020). Determination of Anaerobic Capacity-Reliability and Validity of Sprint Running Tests. *Exercise Science*, 29(2),129-137. doi: 10.15857/ksep.2020.29.2.129
- 124. Weber, S. (2003). Berechnung leistungsbestimmender Parameter der metabolischen Aktivität auf zellulärer Ebene mittels fahrradergometrischer Untersuchungen. Dipl.-Thesis. Cologne: German Sport University Cologne.
- 125. Wittekind, A., & Beneke, R. (2011). Metabolic and performance effects of warm-up intensity on sprint cycling. Scandinavian Journal of Medicine & Science in Sports, 21(6),e201-e207. doi: 10.1111/j.1600-0838.2010.01248.x
- 126. Yang, W., Park, S., Kim, T., Jeon, H., Heine, O., Gehlert, S. (2023). A modified formula using energy system contributions to calculate pure maximal rate of lactate accumulation during a maximal sprint cycling test. *Frontiers in Physiology*, 14. doi: 10.3389/fphys.2023.1147321

- 127. Zouloumian, P., & Freund, H. (1981). Lactate after exercise in man: III. Properties of the compartment model. *European Journal of Applied Physiology and Occupational Physiology*, 46, 149-160. doi: 10.1007/BF00428867
- 128. Zwingmann, L., Hoppstock, M., Wahl, P. (2020). Power profile, physiological characteristics and their correlation in elite canoe polo players. *The Journal of Sports Medicine and Physical Fitness*, 60(9), 1194-1201. doi: 10.23736/s00224707.20.10801-6